# GIBBERELLIC ACID-REGULATED EMBRYO INDUCTION AND GERMINATION IN HEVEA BRASILIENSIS (MUELL. ARG.)

## P. Kumari Jayasree and A. Thulaseedharan

Jayasree, P.K. and Thulaseedharan, A. (2001). Gibberellic acid-regulated embryo induction and germination in *Hevea brasiliensis* (Muell. Arg.) *Indian Journal of Natural Rubber Research*, **14**(2): 106 - 111.

Effect of gibberellic acid (GA<sub>3</sub>) on embryo induction and germination during somatic embryogenesis of  $Hevea\ brasiliensis$  with respect to clone RRII 105 was studied. Immature anthers were inoculated on callus induction medium supplemented with 2,4-dichlorophenoxyacetic acid (2,4-D) and kinetin (KIN) and the induced calli were then transferred to the embryo induction medium. Incorporation of GA<sub>3</sub> up to 2.0 mg/l increased the embryo induction. Germination percentage was significantly enhanced by higher concentrations, however, further plant development was affected by increasing GA<sub>3</sub> levels. A reduction in response to both embryo induction and germination was observed by co-autoclaving of GA<sub>3</sub>.

Key words: Anther culture, Gibberellic acid, Hevea brasiliensis, Somatic embryogenesis.

P. Kumari Jayasree (for correspondence) and A. Thulaseedharan, Rubber Research Institute of India, Kottayam – 686 009, Kerala, India (Email: rrii@vsnl.com).

### **INTRODUCTION**

Challenger and Application .

During in vitro culture, the cells possess or acquire competence for morphogenesis and undergo a permissive pattern of development. Although the success of such pattern of development is affected by various factors, the effect of constituents of medium is very crucial for production of embryo from somatic cells (Ammirato, 1986). In Hevea, somatic embryogenesis using different explants has been reported from China, Malaysia, France and India (Wang et al., 1980; Wan et al., 1982; Carron and Enjalric, 1985; Asokan et al., 1992; Jayasree et al., 1999). Somatic embryogenesis remains problematic due to low germination and plant conversion rate although enough attention has been focussed on its induction phase (Linossier et al., 1997).

Gibberellins (GA) are known to regulate many aspects of growth and development of plants (Hooley, 1994). Gibberellic acid (GA<sub>3</sub>) is a potent growth regulator influencing embryo induction and

(8.00)(1.50

germination. Although conflicting reports are existing for the influence of  $GA_3$  on embryo induction and germination in many crops, a comprehensive study in *Hevea* is lacking. Therefore, it was of interest to investigate the influence of  $GA_3$  on somatic embryogenesis in *Hevea*.

#### **MATERIALS AND METHODS**

Floral buds were collected from *Hevea brasiliensis*, clone RRII 105. After ascertaining the developmental stage, buds at the diploid stage were selected and surface-sterilized with 0.5 per cent hypochlorite solution for 5 min and then washed with sterile distilled water (Jayasree *et al.*, 1999). Immature anthers were dissected out and cultured on modified callus induction medium (Murashige and Skoogs, 1962) containing 2,4-D (2.0 mg/l), KIN (0.5 mg/l) and sucrose (3%). Cultures were incubated under darkness at  $25 \pm 2^{\circ}$ C. Calli induced were then subcultured on embryo induction medium supplemented with glutamine (200

The state of the s

mg/l), casein hydrolysate (400 mg/l),  $\alpha$ -napthaleneacetic acid (NAA) (0.2 mg/l), KIN (0.7 mg/l) and sucrose (7%).

## Effect of GA, on embryo induction

Filter sterilized stock solutions (100 mg/l) of  $GA_3$  at final concentrations of 0.0-10 mg/l with an increment of 1.0 mg/l were added to the above mentioned basal medium after autoclaving. Calli were subcultured on the  $GA_3$  supplemented media and maintained in darkness at  $25 \pm 2^{\circ}\text{C}$ . Efficiency of embryogenesis was calculated on the basis of embryos produced after seven weeks of subculture.

## Influence of GA<sub>3</sub> on embryo germination

Mature embryos which originated from medium containing 2.0 mg/l GA, were transferred to plant regeneration medium containing 3 per cent sucrose and 0.2 per cent activated charcoal (Jayasree et al., 1999). The effect of exogeneous GA, on germination was tested by adding filter-sterilized GA, at 0.0, 1.0, 2.0, 3.0, 4.0 and 5.0 mg/l to the autoclaved medium. Cultures were incubated at  $25 \pm 2^{\circ}$ C under 16 h photoperiod (40 mE/m<sup>2</sup>/s). Efficiency of germination was calculated by scoring the greening of cotyledon, formation of the primary root and shoot followed by the emergence of a leaf. In order to study the effect of GA<sub>3</sub> on coautoclaving with the medium, in separate experiments  $GA_3$  (1.0, 2.0 and 3.0 mg/l) was added to the medium before autoclaving.

All media were adjusted to pH 5.6, gelled with 0.25 per cent gelrite and autoclaved for 10 min at 120°C. The experiments were repeated thrice with 3 replications for each treatment and data were subjected to analysis of variance. Regenerated plants were transplanted to small polybags initially and then to larger polybags (35 x 65 cm). Hardened plants were maintained under shade till field-planting.

#### **RESULTS AND DISCUSSION**

## Effect of GA, on embryo induction

Calli were induced on callus induction medium containing 2,4-D and KIN after 35 days of culture. Upon subculturing these calli to embryo induction medium, discolouration of callus was noticed initially followed by production of friable embryogenic calli and emergence of embryoids (Fig. 1).

Table 1. Influence of gibberellic acid on embryo induction

GA <sub>3</sub> (mg/l)	Mean number of embryos				
	Normal	Abnormal	Total		
0	24	0	24		
1	31	0	31		
2	46	2	48		
3.	27	4	31		
4	18	5	23		
5	11	5	16		
6	5	9	14		
7	3	6	9		
8	1	4	٠ 5		
9	0	2	2		
10	. 0	0	0		
VR	82.74**	10.90**	76.17**		
CD (P=0.05)	4.95	2.55	4.92		

The present study revealed the stimulatory effect of GA<sub>3</sub> at lower levels (Table 1). Though embryo induction occurred even in the absence of GA<sub>3</sub>, it was higher at 1.0 mg/l and maximum at 2.0 mg/l (Fig. 2). A progressive reduction was noticed

Table 2. Effect of gibberellic acid on germination of

embryo				
GA <sub>3</sub> (mg/l)	Cotyledon greening	Root formation	Embryo germination	
	(%)	(%)	(%)	
0	100	100	29.22	
1	100	100	3 <u>9.</u> 56	
2	100	100	41.78	
3	100	100	46.56	
4	100	100	50.22	
5	100	100	52.33	
VR			77.16**	
CD (P=0.05)			2.66	
		_		

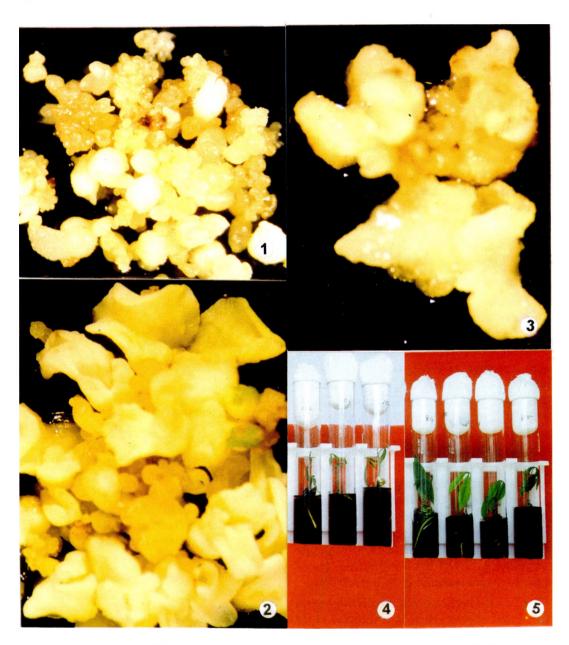


Fig.1-5. Embryogenesis and plant conversion in *Hevea* : 1. Emerging of embryoids from friable callus; 2. Differentiation of various stages of embryos; 3. Embryos with morphological abnormality; 4. Germinated somatic embryos; 5. Fully developed plants.

above 2.0 mg/l with cessation of embryogenesis at 10.0 mg/l. Morphological abnormality (Fig. 3) was frequently noticed in all treatments above 1.0 mg/l. When the

level exceeded 5.0 mg/l, the number of abnormal embryos was higher than normal. For somatic embryogenesis,  $GA_3$  at low levels (0.5 to 2.0mg/l) for sandal wood (Sita

et al., 1979; Sita, 1986) and 1.0 mg/l for fennel petioles and Eryngium foetidum (Hunault and Maatar, 1995; Ignacimuthu et al., 1999) has been found to be optimum. However, enhancement in somatic embryogenesis at 34.6 mg/l for spinach (Xiao and Branchard, 1993) and at 10 mg/l for Rumwex acetosell (Culafic et al., 1987) has also been reported. In contrast, GA<sub>3</sub> appears to suppress embryogenesis in carrot (Tisserat and Murashige, 1977a & b), soybean (Phillips and Collins, 1981), citrus (Kochba et al., 1978) and spinach (Zdravkovic and Neskovic, 1999).

## Effect of GA<sub>3</sub> on germination and plant regeneration

Upon transferring of embryos to hormone-free medium, 29 per cent embryo germination occurred (Jayasree et al., 1999). Germination frequency increased with increasing GA, concentrations (Table 2), however, low level (2.0 - 3.0 mg/l) was found as optimum for both germination and subsequent full plant recovery (Figs. 4 & 5). The embryo germination was more than 50 per cent at 5.0 mg/l which was statistically on par with that at 4.0 mg/l. But full plant development was affected at these concentrations. Though cotyledon greening and root formation was independent of GA, levels, shoot development and leaf appearance were dependent. Retardation of growth at higher concentration of GA, was accompanied by the arresting of shoot elongation and appearance of leaf senescence. In most cases, new leaf development from these plants was delayed or inhibited eventually resulting in the death of the plant. If higher concentrations of GA, was supplied, transfer of germinated embryo to hormone-free medium was found as very essential for further development of plants (data not shown). This suggests that once shoot apex is differentiated, GA, is no longer necessary for further growth. It has been reported that in Eschscholzia californi, addition of GA<sub>3</sub> (3.0 mg/l) though led to an

increase in conversion frequency, resulted in moderate to severe hyperhydricity (Park and Facchini,1999). The growth suppression of germinated embryos of Hevea may also be due to the hyperhydricity. Embryos showing morphological abnormality were also able to germinate in GA, containing medium. GA, has been reported to enhance germination of somatic embryos in Runnex acetosella (Culafic et al., 1987), in grapevine (Mullins and Srinivasan, 1976; Rajasekaran and Mullins, 1979) and in Panax ginseng (Chang and Hussing, 1980; Choi et al., 1999). Although the exact mechanism is not fully clear, the ultrastructural studies carried out by Choi et al. (1999), in ginseng, showed that somatic embryos developed in vitro may be dormant after maturing and thus required a dormancy-breaking treatment.

GA, is a heat sensitive component that can be partly destroyed during autoclaving (Henderson, 1960). In the present study, no significant difference was noticed between addition of filter-sterilized GA, to the autoclaved medium and co-autoclaving of GA, for the induction of embryogenesis. Although reduction in the embryo induction was observed at lower concentrations, a slight enhancement was noticed with autoclaved GA, at 3.0 mg/l (Table 3). However, germination was lower with autoclaved GA, at all concentrations. The marginal reduction in both embryo induction and germination in autoclaved medium may be due to the partial degradation of GA during autoclaving. Hunault and Maatar (1995) also observed that autoclaved GA, was

Table 3. Effect of methods of sterilization of GA,

GA.	Embryo induction (Mean)		Embryo germination (%)	
GA <sub>3</sub>	Filter-	Autoclaved		Attoclaved
	sterilized	:	sterilized	
1.0	31.89	30.89	40.11	35.56
2.0	48.22	45.00	42.56	40.33
3.0	32.22	36.44	47.22	44.56
CD (P=0.05)	1.85	NS	1.96	1.60
CD (interaction	) 2.61	NS		

The state of the s

as effective as filter-sterilized, for somatic embryogenesis of fennel.

In conclusion, lower levels of GA<sub>3</sub> was observed to foster induction and germination of somatic embryos and subsequent development of full plants. Slight reduction in response was observed for both induction and germination of embryos by autoclaving GA<sub>3</sub>.

#### REFERENCES

- Ammirato, P.V. (1986). Control and expression of morphogenesis in culture. In: Plant Tissue Culture and its Agricultural Applications. (Eds. L.H. Wilthers and P.G. Anderson). Butterworth, London, pp. 23-46.
- Asokan, M.P., Jayasree, P. and Sushamakumari, S. (1992).
  Plant regeneration by somatic embryogenesis of rubber tree (*Hevea brasiliensis*). *International Natural Rubber Conference*, 5-8 February 1992, Bangalore, India, p. 49.
- Carron, M.P. and Enjalric, F. (1985). Embryogenese somatique a partir du tegument iinterne de la graine d' *Hevea brasiliensis* Muell-Arg. CR Acad Sci, Paris, 300 (111) 17: 653-658 (English summary).
- Chang, W.C. and Hussing, Y.I. (1980). Plant regeneration through somatic embryogenesis in root derived callus of ginseng (*Panax ginseng C.A. Meger*). Theoretical and Applied Genetics, **57**: 135-138.
- Choi, Y.E., Yang, E.S., Yoonk, K. and Choi, T. (1999). High efficiency plant production via direct somatic embryogenesis from preplasmolysed cotyledons of *Panax ginseng* and possible dormancy of somatic embryos. *Plant Cell Reports*, 18: 493-499.
- Culafic, L., Budmr, S., Vujicic, R. and Neskovic, M. (1987). Induction of somatic embryogenesis and embryo development in Runnex acetosella L. Plant Cell, Tissue and Organ Culture, 11: 133-139.
- Henderson, J.H.M. (1960). Influence of hydrogen ion concentration and autoclaving on gibberellin. *Nature*, **185**: 628-629.
- Hooley, R. (1994). Gibberellins: Perception, transduction and responses. Plant Molecular Biology, 26: 1529-1555.
- Hunault, G. and Maatar, A. (1995). Enhancement of somatic embryogenesis frequency by gibberellic acid in fennel. *Plant Cell, Tissue and Organ Culture*, 41:171-176.
- Ignacimuthu, S., Arociasamy, S., Antonysamy, M. and Ravichandran, P. (1999). Plant regeneration

#### **ACKNOWLEDGEMENT**

The authors wish to thank Dr. N.M. Mathew, Director, Rubber Research Institute of India for encouragement and support for undertaking this study. The help rendered by Mr. Ramesh B. Nair, Assistant Director (Statistics) for statistical analysis of the data is gratefully acknowledged.

- through somatic embryogenesis from mature leaf explants of Eryngium foetidum, a condiment. Plant Cell, Tissue and Organ Culture, 56: 131-137.
- Kochba, J., Spiegel-Roy, P., Neumann, H. and Saad, S. (1978). Stimulation of embryogenesis in Citrus Callus by ABA, Ethephon, CCC & Alar and its suppression by GA. Zpflanzenphysiol, 89: 427-432.
- Jayasree, P.K., Asokan, M.P., Sobha, S., Sankaraiammal, L., Rekha, K., Kala, R.G., Jayasree, R. and Thulaseedharan, A. (1999). Somatic embryogenesis and plant regeneration from immature anthers of Hevea brasiliensis (Muell. Arg.). Current Science, 76(9): 1242-1245.
- Linossier, L., Veisseire, P., Cailloux, F. and Coudret, A. (1997). Effect of abscisic acid and high concentrations of PEG on *Hevea brasiliensis* somatic embryos development. *Plant Science*, **124**: 183-191.
- Murashige, T. and Skoogs, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiologia Plantarum*, **15**: 473-497.
- Mullins, M.G. and Srinivasan, C. (1976). Somatic embryos and plantlets from an ancient clone of the grapevine (CV Cabernet Sauviignon) by apomixes in vitro. Journal of Experimental Botany, 27: 1022-1030.
- Park, S.U. and Facchini, P.J. (1999). High efficiency somatic embryogenesis and plant regeneration in California poppy, Eschscholzia california cham. Plant Cell Reports, 19: 421-426.
- Phillips, G.C. and Collins, G.B. (1981). Induction and development of somatic embryos from cell suspension cultures of soyabean. *Plant Cell, Tissue and Organ Culture*, 1:123-129.
- Rajasekaran, K. and Mullins, M.G. (1979). Embryos and plantlet from cultured anthers of hybrid grapevines. Journal of Experimental Botany, 30:399-407.
- Sita, G.L. (1986). Sandalwood (Santhalum album L.). In: Biotechnology in Agriculture and Forestry (Ed. Y.P.S. Bajaj). Trees, Springer Verlag, Berlin, Herde Pberg, pp. 1: 363-374.

- Sita, G.L., Ram, N.V.R. and Vaidyanathan, C.S. (1979).
  Differentiation of embryoids and plantlets from short callus of sandalwood. *Plant Science Letters*, 15: 265-270.
- Tisserat, B. and Murashige, T. (1997a). Repression of asexual embryogenesis *in vitro* by some plant growths. *In vitro*, 13: 779-805.
- Tisserat, B. and Murashige, T. (1997b). Effects of ethephon, Ethylene and 2,4-dichlorophenoxyacetic acid on asexual embryogenesis in vitro. *Plant Physiology*, **60**: 437-439.
- Wan, A.R., Ghandimathi, H., Rohini, O. and Paranjothy, K. (1982). Recent developments in tissue culture of *Hevea*. In: Tissue Culture of Economically

- Important Plants (Ed. A.N. Rao). Costed, Singapore, pp. 152-158.
- Wang, Z., Zeng, X., Chen, C., Wu, H.L., Fan, G. and Lu, W. (1980). Induction of rubber plantlets from of *Hevea brasiliensis* Muell. Arg. in vitro. *Chinese Journal of Tropical Crops*, 1(1): 25-26.
- Xiao, X.G. and Branchard, M. (1993). Embryogenesis and plant regeneration of spinach (Spinacia oleracea L.) from hypocotyl segments. Plant Cell Reports, 13: 69-71.
- Zdravkovic, K. and Neskovic, M. (1999). Induction and development of somatic embryogenesis from spinach (Spinacia oleracea) leaf segments. Plant Cell, Tissue and Organ Culture, 50: 109-114.