

Agro's Ann. Rev. Pl. Physiol. (B & A). Vol. 2. S.S. Purohit (Ed.). pp 281-317.  
© 1996-97, Agro Botanica, Bikaner, India

## GROWTH AND DEVELOPMENT OF PLANTS IN A CO<sub>2</sub> ENRICHED WORLD

A. S. DEVAKUMAR AND JAMES JACOB

Plant Physiology Division, Rubber Research Institute of India,  
Kottayam - 686 009, Kerala, India

GROWTH AND DEVELOPMENT OF PLANTS IN A CO <sub>2</sub> ENRICHED WORLD.....	281
PHOTOSYNTHESIS.....	283
STOMATA AND TRANSPIRATION.....	289
RESPIRATION.....	290
PRODUCTION AND PARTITIONING OF DRY MATTER BETWEEN ORGANS.....	291
Leaves.....	292
Roots.....	293
Reproductive Development.....	295
Economic Yield.....	295
TISSUE COMPOSITION.....	296
SENESCENCE.....	297
INTERACTION OF ATMOSPHERIC CO <sub>2</sub> WITH ENVIRONMENTAL FACTORS.....	298
Temperature.....	298
Light.....	299
Nutrients.....	300
Moisture Stress.....	300
ELEVATED CO <sub>2</sub> AND ECOSYSTEM PROCESSES.....	301
Biomass.....	301
Insects and Fungi.....	301

INFLUENCE OF ELEVATED CO <sub>2</sub> ON PLANT-PLANT	
INTERACTION.....	303
CONCLUSIONS.....	304
REFERENCES .....	305

The never ending wants of human race for a better life has forced to discover as well as exploit the natural resources. The material comforts that man has realized so far are highly energy dependant. Globally, the lion share of the human energy needs is met by fossil fuels which leads to emission of CO<sub>2</sub> apart from CO, SO<sub>2</sub> and N<sub>2</sub>O into the atmosphere. There is an environmental cost to be paid for the energy that we consume.

Industrial revolution and urbanization has led to clearing of large areas of forests. Forests are reported to contribute 70% of the terrestrial carbon fixation, there by making them a major sink for atmospheric CO<sub>2</sub> (Waring and Schlesinger, 1985). Because of deforestation and burning of fossil fuel an imbalance between the sources and sinks for CO<sub>2</sub> occurs, leading to the build up of CO<sub>2</sub> in the atmosphere. Estimates of CO<sub>2</sub> concentrations for a period of 200 years between 1750 - 1950 have indicated the rate at which CO<sub>2</sub> was building up in the atmosphere (Neftal *et al*, 1985). Between 1750 - 1850 a small increase in the atmospheric CO<sub>2</sub> was estimated to occur from 280 ppm to 285-290 ppm at the rate of 0.05-0.1 ppm year<sup>-1</sup>. However, between 1850 - 1950, due to industrial revolution accompanied by population pressure, the concentration increased from 290 to 315 ppm which accounts for an average rate of increase of 0.35 ppm year<sup>-1</sup>. In the last forty five years or so it has increased to more than 350 ppm, the average rate of increase being 0.83 ppm year<sup>-1</sup>. Currently the CO<sub>2</sub> concentration is increasing at a rate of 1.8 ppm year<sup>-1</sup> (John and Lloyd, 1992). Unless radical steps are taken to contain fossil fuel burning, the atmospheric CO<sub>2</sub> concentration may reach 50% above the pre-industrial concentrations in the next 20 years. Photosynthesis being the only process through which plants absorb atmospheric CO<sub>2</sub> it plays an important role in the carbon cycle. There is growing interest to know how plants and ecosystems would respond to continued increases in atmospheric CO<sub>2</sub> concentrations.

In the past two decades, quite a good amount of research has gone in to the analysis of plant response to higher concentrations of CO<sub>2</sub>. Though there is much to be understood, it is evident from the majority of these studies that there will be increased growth rates, especially in plants with the C<sub>3</sub> type of photosynthetic metabolism. The primary plant process that is affected due to higher concentrations of CO<sub>2</sub> is photosynthesis. It is well established that photosynthetic rates of C<sub>3</sub> plants are limited by the existing atmospheric CO<sub>2</sub> levels (Farquhar, 1980; Sharkey, 1985) and therefore, photosynthetic rates in these plants are expected to increase under elevated CO<sub>2</sub> concentrations. Plant stomatal dynamics is another character which plays a key role in allowing CO<sub>2</sub> to enter the plant system and at the same time transpire moisture from the plant body, hence regulating the water use efficiency. How these two processes are co-ordinated under elevated CO<sub>2</sub> becomes important. Respiration is the major catabolic process through which plant loses nearly 50% of the carbon fixed in the process of photosynthesis. Therefore, regulation of respiration under elevated CO<sub>2</sub> levels can alter the carbon balance of plants. Allocation of carbon between various plant organs, reproductive development and senescence are other important issues that are altered due to growth in a CO<sub>2</sub> enriched atmosphere. Response of individual plants to elevated CO<sub>2</sub> will affect various ecosystem processes in the long term. This review is an attempt to cover the responses of plants to elevated CO<sub>2</sub> at different levels of organization of the ecosystem.

### PHOTOSYNTHESIS

Plants respond to atmospheric CO<sub>2</sub> through photosynthesis. The primary carboxylase enzyme, ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) present in C<sub>3</sub> plants is sensitive to atmospheric CO<sub>2</sub> concentration (Bowes, 1993). This enzyme has the dual role of carboxylase and oxygenase. At the prevailing CO<sub>2</sub> concentration, the carboxylase activity is not saturated and it is estimated that almost 30% or even more of the carbon assimilated by the plant is released back into the atmosphere through the processes of photorespiration resulting from the oxygenase activity of this enzyme. An increase in the atmospheric CO<sub>2</sub> concentration will decrease the oxygenase activity and increase the carboxylase activity resulting in increased photosynthesis in a CO<sub>2</sub> enriched atmosphere. A doubling of atmospheric CO<sub>2</sub> concentration from the present level will inhibit

photorespiration by 50% (Sharkey 1988). The immediate and marked response of plants to elevated  $\text{CO}_2$  owes to the biochemical properties of Rubisco (Long, 1991).

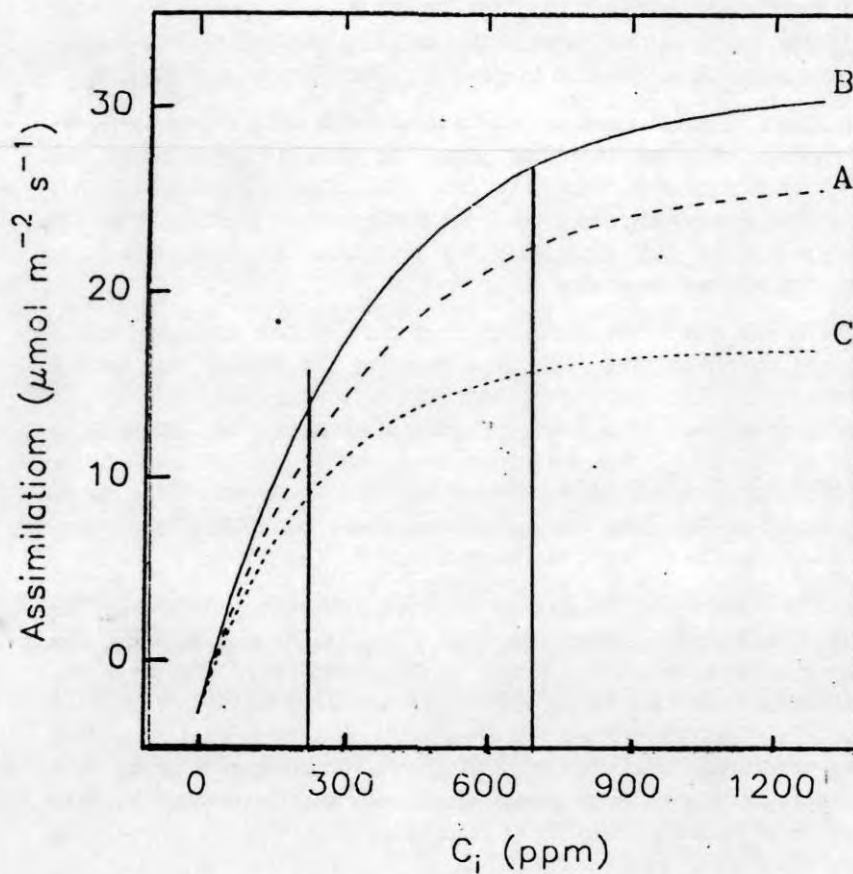


Figure 1. Three hypothetical curves to show various types of photosynthetic acclimation responses. Curve A represents a plant grown in normal ambient  $\text{CO}_2$ . Curves B and C represent plants grown at elevated  $\text{CO}_2$  concentration; B shows a positive acclimation and C shows a negative acclimation. The vertical lines indicate the  $C_i$  corresponding to a  $C_a$  of 300 ppm and 700 ppm  $\text{CO}_2$ .

Table 1: The following studies have shown significantly higher photosynthetic rates in plants grown at elevated CO<sub>2</sub> than those grown at normal ambient CO<sub>2</sub> when compared at their respective growth CO<sub>2</sub> concentration

Species	Reference
<i>Scirpus olneyi</i>	Jacob et al., 1994
<i>Ranunculus glacialis</i>	Kroner and Diemer, 1994
<i>Loblolly pine</i>	Tissue et al., 1993
White oak, yellow poplar	Gunderson et al., 1993
<i>Populus grandidentata</i>	Curtis and Teir, 1992
White oak, yellow poplar	Wullschlegel et al., 1992
<i>Scirpus olneyi</i>	Arp and Drake, 1991
Soybean	Ziska et al., 1990; Campbell et al., 1988; Clough et al., 1981; Havelka et al., 1984; Cure et al., 1989
Potato	Sage et al., 1989
Cotton	Wong, 1979
(grown with high N)	
Vine	Kriedman et al., 1976
<i>Pinus taeda</i>	Lewis et al., 1996

The following studies have shown lower (or smaller) rates of photosynthetic rates in plants grown at elevated CO<sub>2</sub> than those grown at normal ambient CO<sub>2</sub> when compared at their respective growth CO<sub>2</sub> concentrations:

Species	Reference
<i>Phaseolus vulgaris</i>	von Caemmerer and Farquhar 1984; Sage et al., 1989
<i>Brassica oleracea</i>	Sage et al., 1989
<i>Chenopodium album</i> ,	
Cotton	Thomson and Strain, 1991
Water Hyacinth	Spencer and Bowes, 1986
Cotton (Grown with low N)	Wong, 1979
Tomato	Yelle et al., 1989a; Besford, 1993
<i>Eriophorum vaginatum</i>	Tissue and Oechel 1987
Tobacco	Raper and Peedin 1978
Cucumber	Peet et al., 1986
<i>Eucalyptus</i>	Roden and Ball, 1996a, 1996b
tomato	Van ooster and Besford, 1995

Many studies have shown that photosynthesis will increase with CO<sub>2</sub> enrichment. (see above Table 1 for a summary). In a study with six different C3 species, a significant increase in the photosynthetic rates was found at elevated CO<sub>2</sub> levels (Ziska, *et al.*, 1991). In soybean 1.5 fold increase of assimilation was seen with CO<sub>2</sub> enrichment (Wong 1990). Long and Drake (1992) has reviewed the photosynthetic response to elevated CO<sub>2</sub> in more details and concluded that there is considerable stimulation in photosynthesis with CO<sub>2</sub> enrichment.

However, there are also reports which are contradictory to the above type of plant response. Photosynthetic rates of plants grown in elevated CO<sub>2</sub> can be often lower than the rates of plants in normal ambient CO<sub>2</sub> when measured at the same back ground CO<sub>2</sub> (DeLucia *et al.*, 1985). This is generally termed photosynthetic acclimation.

The relevant question for a future world rich in CO<sub>2</sub> is that whether plants would adjust their photosynthetic response to elevated CO<sub>2</sub> in the long run. Would they undergo acclimation? If so what would be the extent of photosynthetic acclimation? And will the degree of acclimation vary with the plant type? Three hypothetical scenarios of no acclimation and a positive or negative acclimation are demonstrated in Figure 1.

It has been shown that prolonged exposure to elevated CO<sub>2</sub> results in a gradual reduction in the photosynthetic capacity of these plants. The photosynthetic rate at a given background CO<sub>2</sub> level will be less in a plant exposed to elevated CO<sub>2</sub> for long periods of time than a plant grown at the normal ambient CO<sub>2</sub>. This is often reflected in the reduced initial slope of the A/Ci response function (carboxylation efficiency). This reduction in the carboxylation efficiency was found to be related to a reduction in the total soluble protein pool, including Rubisco of the leaves grown at elevated CO<sub>2</sub> (Jacob *et al.*, 1994; Stitt, 1991). However, seldom was the photosynthetic rates of the plants grown and measured at elevated CO<sub>2</sub> less than that of plants grown and measured at normal ambient CO<sub>2</sub>. In other words, at the operational CO<sub>2</sub> levels plants grown at elevated CO<sub>2</sub> did maintain a higher photosynthetic rate than their counter parts grown at normal ambient CO<sub>2</sub> (Stitt, 1991; Long and Drake, 1992).

Indirect measurements of Rubisco protein by binding with <sup>14</sup>C carboxyarabinitol 1-5, Bisphosphate (CABP) indicated that there was loss of Rubisco protein in cabbage grown in elevated CO<sub>2</sub> (Sage *et al.*, 1989). There was an initial increase in photosynthetic rates, followed by



a decrease over days or weeks to a rate similar to that observed in plants grown at ambient CO<sub>2</sub> (Delucia *et al.*, 1985). The decrease in Rubisco content has been attributed to the reallocation of nitrogen to the other more limiting enzyme systems (Woodrow 1994). Because carboxylation is favored through higher CO<sub>2</sub> availability in plants grown at elevated CO<sub>2</sub>, decreased Rubisco content appears to be the major factor leading to photosynthetic acclimation (Sage *et al.*, 1989; Arp *et al.*, 1991).

There are also some reports of a reduction in the photosynthetic rate of elevated CO<sub>2</sub> grown plants below the rates of those grown and measured at normal ambient CO<sub>2</sub> (Diaz *et al.*, 1994). This kind of down regulation is generally observed in pot studies and in annuals. There are few reports of such negative down regulation in photosynthesis in perennial and in field studies.

Thomas and Strain (1991) demonstrated a decline in photosynthetic capacity in potted cotton plants because of the smaller size of the containers in which they were grown. The decline in photosynthesis was rapidly reversible if plants were transplanted to larger pots. According to Cure and Acock (1986) a high metabolic or storage sink activity is required for sustained photosynthetic response to elevated CO<sub>2</sub>. Long and Drake (1992) found out that photosynthetic stimulation to elevated CO<sub>2</sub> was less (23%) in pot studies than field (65%) studies.

The source-sink balance is a major factor in determining the acclimation of photosynthesis to high CO<sub>2</sub>. Variabilities affecting this balance may influence the extent of the reduction of photosynthetic capacity in high CO<sub>2</sub> (Arp, 1991). Arp and Drake (1991) concluded from a long term experiment that after four years of exposure to elevated CO<sub>2</sub>, the photosynthetic capacity of *Scripus olneyi* grown in the field was not decreased. Field grown crops and natural ecosystems, may continue to respond positively to increasing atmospheric levels of CO<sub>2</sub> (Gunderson *et al.*, 1993).

It is proposed that the source sink relationship of the plant plays a key role in the degree of acclimation to elevated CO<sub>2</sub> and this is brought about by alterations in the rates of supply and utilization of photosynthates. In annuals and in plants grown in pots, probably the sink demand for more photosynthates is not high enough to maintain a steady high rate of photosynthesis at elevated CO<sub>2</sub>. This results in accumulation of carbohydrates in the source organs which will down regulate photosynthesis through various feed back regulatory mechanisms.

Accumulation of starch and soluble sugars in plants grown at elevated  $\text{CO}_2$  is commonly observed (Long and Drake, 1992). Starch accumulation can lead to reduced availability of light to chloroplast (Neals and Nicholls, 1978) tie up inorganic phosphate in metabolic intermediaries and even rupture chloroplast (Stitt, 1991; Caveer *et al.*, 1981; Woulf and Strain, 1982).

It has been shown that accumulation of soluble sugars may be involved in down regulating the expression of Rubisco genes and possibly other photosynthetic genes and thus regulate photosynthesis resulting in acclimation of the photosynthetic apparatus (Nie *et al.*, 1995b). A review by Long and Drake (1992) shows that in plants grown in pots, starch and sucrose increased by more than 300% and 150%, respectively. It is possible that organs such as rhizomes, hard wood, stem etc may act as a repository for photoassimilates and thus increase the demand for photoassimilates more in perennials than annuals. It may be noted that a complete negation of the stimulatory effect of elevated  $\text{CO}_2$  on photosynthesis was never seen in perennials or in field grown plants, but was observed only in a few cases of pot culture experiments with annual crop plants.

Diurnal changes in the stimulation in photosynthesis to elevated  $\text{CO}_2$  was affected by end product accumulation and faster movement of photoassimilates from the leaves to the sink was necessary to maintain the continued stimulation in photosynthesis (Zhang and Nobel 1996). Nie *et al.*, (1995b) have reported increased leaf soluble carbohydrates concentration and decreased mRNA transcripts for genes coding for some photosynthetic enzymes and in particular, *rbcS* and *rbcL* coding for Rubisco small unit and large subunits, respectively. Feeding glucose to leaves repressed the expression of many Calvin cycle genes (Jones *et al.*, 1996; Sheen 1990). But Riviere-Rolland *et al.*, (1996) reported that Rubisco is not down regulated beyond a threshold supply of N even if soluble carbohydrates accumulate. Activities and transcript levels of Rubisco and carbonic anhydrase were less in plants grown at elevated  $\text{CO}_2$  (Majeau and Coleman, 1996). Exposed leaves did not down regulate Rubisco, but shaded leaves did in response to elevated  $\text{CO}_2$  (Nie *et al.*, 1995b). It has been suggested that Rubisco is down regulated to tune the rate of photosynthesis in equilibrium with the sink activity of the plant (Stitt, 1991). The capacity for sucrose synthesis may also limit the plants' ability to respond to elevated  $\text{CO}_2$ . Flaveria plants with less fructose-1,6-bisphosphatase had less response to elevated  $\text{CO}_2$  (Micallef



*et al.*, 1996). Acclimation of photosynthesis to elevated CO<sub>2</sub> appears to be a classical case of source sink dynamics mediated through end product induced down regulation of events at the molecular and physiological levels.

### STOMATA AND TRANSPIRATION

Irrespective of the degree of acclimation in photosynthesis, stomatal conductance and transpiration rates are less in plants exposed to elevated CO<sub>2</sub>. This is largely true in short and long term experiments. This results in higher water use efficiency in plants grown at elevated CO<sub>2</sub>. If stomata behave in such a way to keep the Ci/Ca ratio constant at nearly 0.7, it is estimated that the stomatal conductance should be decreased by 40% when the CO<sub>2</sub> concentration is increased from 350 to 700 ppm (Long and Drake 1992). However, what is observed in terms of reduction in stomatal conductance and increase in WUE at the single leaf level may not be fully translated into canopy levels, because, plants grown at elevated CO<sub>2</sub> tend to have more leaf area and biomass and thus end up losing more water than what is predicted from single leaf measurements. But, for comparable leaf area and biomass there is definitely less requirement of water in plants grown at elevated CO<sub>2</sub> than normal ambient CO<sub>2</sub>.

A reduction in the transpiration rates has immense implications to the plants, soil and atmosphere. Less transpiration means that the leaf tends to remain slightly warmer. This will affect the leaf metabolism including the response to elevated CO<sub>2</sub>. The activity of many enzymes including Rubisco are temperature sensitive. Photosynthesis is highly sensitive to temperature (Long, 1991). It may be noted that Rubisco may become more of an oxygenase and less of a carboxylase as temperature increases, because temperature alters the CO<sub>2</sub>/O<sub>2</sub> specificity of Rubisco and the solubilities of CO<sub>2</sub> and O<sub>2</sub> in water (Jordan and Ogren, 1984). Growth and development of organs are extremely temperature dependent. Reduced water loss from the leaf will improve the water status of the leaves and the plant in general. This helps to conserve soil moisture and thus helps in managing drought to some extent. Reduced transpiration will alter the microclimate, particularly the relative humidity in the immediate environment of plants which will have implications for other living organisms sharing the same ecosystem with the plants. Reduced stomatal conductance may eventually result in less canopy conductance leading to substantial reduction in the transpiration

rates of plants in large geographical areas and thus affecting the local weather markedly. eg. increased atmospheric temperature or reduced precipitation (Field *et al.*, 1995).

Kimball and Idso (1983) have shown that doubling CO<sub>2</sub> levels decreased transpiration by 30% each in soybean and cotton and 10% in wheat, but by 45% in maize. Allen, (1990) showed 23% decrease in transpiration in maize. So the range is large and variable. There was 30% decrease in transpiration in Eucalyptus at 680 ppm of CO<sub>2</sub> and at saturating light intensity compared to normal ambient CO<sub>2</sub> (Wong and Dunin, 1987). Jones *et al.*, (1985) reported a significant decrease in transpiration rate of soybean plants at elevated CO<sub>2</sub>. Wong and Dunin (1987) noticed 80% reduction in transpiration ratio which was due to a 50% increase in photosynthetic rates and a 30% reduction in transpiration. Elevated CO<sub>2</sub> induced increase in WUE has been reported in many plant species (Oberbauer *et al.*, 1985; Picon *et al.*, 1996). There was 63% improvement in the canopy WUE and a three fold increase in single leaf WUE at elevated CO<sub>2</sub> levels in white clover (Nijs *et al.*, 1989). Data obtained for tropical plants indicate that the WUE increases substantially under elevated CO<sub>2</sub> conditions (Ziska *et al.*, 1991). This higher WUE is attributable to the combined effect of reduced transpiration along with increased photosynthetic rates. This trend was noticed in both C3 and C4 species (Morrison, 1985). For tropical tree species, Reekie and Bazzaz, (1989) report a tendency towards a lower rate of water loss, with increasing CO<sub>2</sub> concentrations.

## RESPIRATION

One would expect that the increased supply of photosynthates would stimulate respiration in plants grown at elevated CO<sub>2</sub>, because photosynthates are the basic substrates which when oxidized during respiration provide the carbon skeleton and the energy for growth and maintenance of the plant. Respiration has been shown to increase in response to short term increases in the concentration of exogenously applied sugars (Azcon-Bieto 1983). But there is considerable reduction in the respiration rates of plants grown at elevated CO<sub>2</sub> (Amthor, 1991; Bunce, 1990) in spite of increased availability of sugars. This decrease is particularly observed in mature tissues exposed to elevated CO<sub>2</sub> (Bunce, 1990). In young tissues, increased photosynthetic rates stimulated respiration rates under elevated CO<sub>2</sub> (Hrubec *et al.*, 1985; Poorter *et al.*, 1988).

Effects of elevated CO<sub>2</sub> on respiration can be classified into two categories. The first one is the direct effect of elevated CO<sub>2</sub>. This is the immediate reduction in the respiratory rate when plants are exposed to elevated CO<sub>2</sub> in the short term and this is often completely reversible. The second is a long term residual effect which is related to changes in the tissue biochemistry and it is less reversible. In this case the respiratory rates of plants grown at elevated CO<sub>2</sub> will be less than the rates of plants grown at normal ambient CO<sub>2</sub> when both set of plants are measured at the normal ambient CO<sub>2</sub> level. This type of response is related to the long term acclimatrory changes that have occurred to the biochemical composition of the respiring tissue (Farrar and Williams, 1991). The most important of them are the changes in the total soluble protein content of the tissues leading to a significant reduction in the tissue N content (Bowes, 1993). A reduction in the tissue N content may explain only part of the reduction in the respiration at elevated CO<sub>2</sub> (Azcon-Bieto *et al.*, 1994). They have found that reduction in the respiration at elevated CO<sub>2</sub> was correlated with reduction in the activities of mitochondrial enzymes, particularly Cytochrome c oxidase. Elevated CO<sub>2</sub> has a direct effect on the mitochondrial respiration (cytochrome c oxidase pathway) and not on the cyanide resistant alternative pathway (Gonzalez-Meler, 1996).

Inhibition in the respiratory rates of plants may contribute substantially to the over all biomass production of plants grown at elevated CO<sub>2</sub> over and above what might be expected from the enhanced photosynthetic rates (Reuvei & Gale 1985).

### PRODUCTION AND PARTITIONING OF DRY MATTER BETWEEN ORGANS

Reasonably good number of plant species were studied for their response to rising levels of CO<sub>2</sub> concentration. To quote a few, CO<sub>2</sub> enrichment increased dry matter (DM) production in cereal crops like wheat (Gifford, 1979, Schonfeld *et al.*, 1989; Chaudhuri *et al.*, 1990), rice (Baker *et al.*, 1990), vegetable crops like cucumber, cabbage and carrot (Slack and Hand, 1985; Heij *et al.*, 1984; Wheeler *et al.*, 1994) and fruit crops like oranges and tomatoes (Downtown *et al.*, 1987; Calvert, 1975; Idso & Kimball, 1994), tuber crops like cassava, sweet potato and potato (Imai *et al.*, 1984; Biswas and Hileman, 1985; Ku and Edwards, 1977) and fibre crops like cotton (Hendrix *et al.*, 1994). Extensive work on oil seeds like soybean also indicated the increased

production of DM at elevated CO<sub>2</sub> concentration (Havelka, *et al.*, 1984a). Not just the food crops or crops of commercial importance that would gain from enriched CO<sub>2</sub> concentrations; even weeds like water hyacinth and *Echinochloa crusgalli* accelerated their growth rates and produced more DM under elevated than normal ambient CO<sub>2</sub> (Poorter *et al.*, 1988; Potvin and Strain, 1985). Increased biomass in plants grown at elevated CO<sub>2</sub> has been reported by Schenk *et al.*, 1995; Rogers *et al.*, 1996a; 1996b; Nobel *et al.*, 1996; Hunt *et al.*, 1996; Roden and Ball, 1996b).

### Leaves

Elevated CO<sub>2</sub> levels increase the leaf area (LA) either by increasing the leaf number or by higher expansion rates. Such an increase in LA under elevated CO<sub>2</sub> condition was recorded in soybean (Rogers *et al.*, 1983) and many tree species (Devakumar, 1994). Increase in LA is largely due to more extensive branching in dicotyledonous plants (Rogers *et al.*, 1984) and tillering in grasses (Sionit *et al.*, 1981). Increase in LA is also attributed to increase in expansion rates and a small increase in maximum leaf size (Cure *et al.*, 1989). However, Biswas and Hileman (1985) found increase in LA in sweet potato was primarily due to an increase in branching and greater leaf size. Increase in total number of main stem, leaf nodes and the area of the main stem trifoliate leaves in soybean is shown by Miglietta *et al.*, (1993). Marginal increase in the rate of leaf initiation due to CO<sub>2</sub> enrichment was seen in soybean (Baker *et al.*, 1989; Cure *et al.*, 1989), winter wheat (Sehonfeld *et al.*, 1989) and weed species *Pueraria lobata* (Sesak and Strain, 1989).

Specific leaf weight (SLW) increased in response to elevated CO<sub>2</sub> in soybean (Leadley and Reynolds, 1988; Leith *et al.*, 1986; Rogers *et al.*, 1983) and sweet potato (Biswas and Hileman, 1985). The increase in SLW was presumably due to the increase in starch content (Huber *et al.*, 1984). Increase in leaf thickness was also attributed to the increased number of palisade cells as seen in soybean (Thomas and Harvey, 1983). On the other hand, maize plants grown in elevated CO<sub>2</sub> did not show any remarkable increase in SLW (Rogers *et al.*, 1983). Elevated CO<sub>2</sub> seemed to have little effect on increasing palisade cells in maize (Thomas and Harvey, 1983).

### Roots

In various field and pot experiments with different crops root growth was found to increase when plants are exposed to elevated CO<sub>2</sub> concentrations (Del castillo *et al.*, 1989). In soybean plants root length was found to increase. The elongation rate of individual root axis was not affected, but there was a significant increase in the number of actively growing roots (Del castillo *et al.*, 1989).

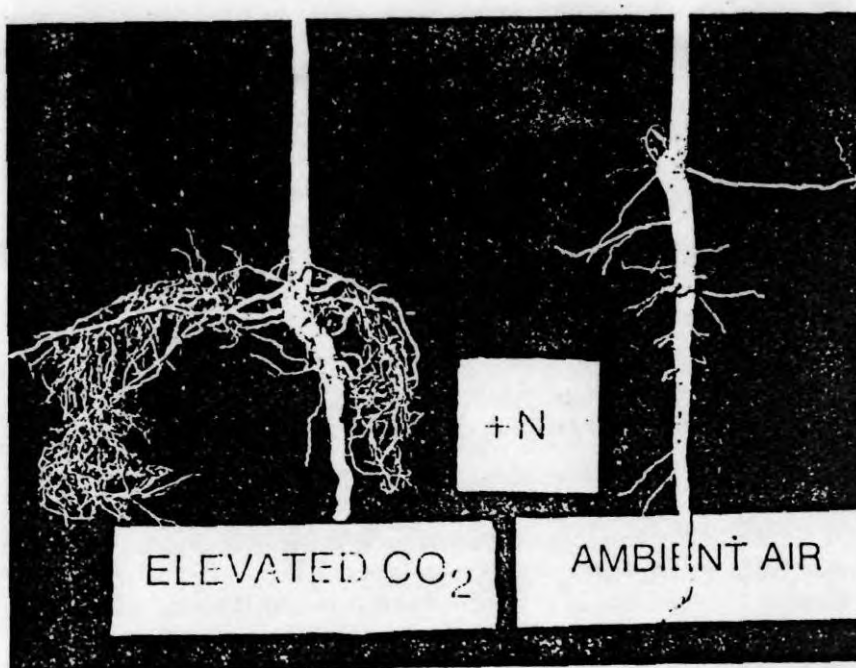


Fig. 2. Allocation of biomass to root growth in seedlings of *Hevea brasiliensis* grown under elevated CO<sub>2</sub> concentration and at ambient air. Seedlings were grown with normal dose of nitrogen (+N) fertilizer in the nursery bed.



CO<sub>2</sub> enrichment was found to increase both root biomass at all depths and maximum depth reached by the roots (Baker *et al.*, 1990, Chaudhuri *et al.*, 1990). Similar type of response was seen in sour oranges (Idso and Kimball, 1992). In native vegetation also elevated CO<sub>2</sub> increased root production (Day *et al.*, 1996). Under moisture stress conditions, root growth was found to increase when higher CO<sub>2</sub> concentration was provided indicating that CO<sub>2</sub> can compensate for decreased root growth under drought (Del Castillo *et al.*, 1989). However, the root dry matter of sweet potato increased more with elevated CO<sub>2</sub> in well watered than in droughted plants (Bhattacharaya *et al.*, 1990). In one of our studies where seedlings of *Hevea brasiliensis* were grown in the nursery beds without any restriction of soil volume, and with normal dose of N, there was a substantial increase in root growth of plants grown under elevated CO<sub>2</sub> (Fig. 2) leading to high root to shoot ratio (Devakumar *et al.*, 1996).

Varying trends were evident from control environment studies on the effect of elevated CO<sub>2</sub> on the distribution of dry matter between organs. For example, no significant effect of elevated CO<sub>2</sub> was seen on the harvest indices and root:shoot ratios (Cure and Acock, 1986). However, in crops where tubers form the bigger proportion of DM, the response was different.

In field grown sweet potato increased diversion of total dry matter to tubers was observed at higher concentrations of CO<sub>2</sub> (Biswas and

Hileman, 1985; Bhattacharaya *et al.*, 1990). Similar results were found in carrot and radish (Idso *et al.*, 1988), but there was little effect of elevated CO<sub>2</sub> on the root:shoot ratio in cotton and soybean (Idso *et al.*, 1988; Radin *et al.*, 1987). An increase in harvest index due to elevated CO<sub>2</sub> was observed in cowpea (Biswas and Hileman, 1985), but there was no effect on the winter wheat (Havelka *et al.*, 1984b). There were no significant changes in the partitioning of dry matter between leaves, stems or pods due to elevated CO<sub>2</sub> in one study on soybean (Ackerson *et al.*, 1984), although there was a slight decline in harvest index with increasing CO<sub>2</sub> in another (Rogers *et al.*, 1986). The pattern of biomass distribution between various organs may be influenced by experimental conditions and availability of nutrients.

### Reproductive Development

Elevated CO<sub>2</sub> influences the reproductive biology of plants. Floral initiation was observed to be 1-3 days earlier in wheat and sunflower due to higher CO<sub>2</sub> concentration, while the rate of primordial initiation was reduced (Marc and Gifford, 1984). Similarly, flowering was 1-3 days earlier in faba bean, wheat and leucerne (Goudrian and de Rutier, 1983). However, no effect of elevated CO<sub>2</sub> concentration on the timing of reproductive stages was found in soybean (Havelka *et al.*, 1984a; Rogers *et al.*, 1986). Lawlor and Mitchell (1990) are of the opinion that these small changes in reproductive stages with CO<sub>2</sub> are not a direct influence of CO<sub>2</sub> concentration on development.

### Economic Yield

Apart from increasing the total dry matter, CO<sub>2</sub> enrichment has also been shown to increase the economic yield. This increase in yield due to CO<sub>2</sub> enrichment is often attributed to an increase in the number of structures rather than in their mean size. In soybean, yield increased almost entirely as a result of more number of seeds (Ackerson *et al.*, 1984; Havelka *et al.*, 1984a; Rogers *et al.*, 1983). However, the number of seeds per pod decreased as a result of CO<sub>2</sub> enrichment (Ackerson *et al.*, 1984). The number of seeds per pod decreased at elevated CO<sub>2</sub> with a concomitant increase in the number of pods in *Phaseolus vulgaris* (Gustafson, 1984).

Interestingly, in some cases, CO<sub>2</sub> enrichment did not affect the mean seed weight. The number of panicles per plant was almost entirely responsible for a 47% increase in grain yield due to a doubling of CO<sub>2</sub> concentration in rice (Baker *et al.*, 1990) and in wheat (Havelka *et al.*, 1984b). However, Krenzer and Moss (1975) concluded that increase in spring wheat yield was due to an increase in grain weight. Sweet potato yields increased as a result of more tubers rather than mass per tuber (Biswas and Hileman, 1985). The effect of CO<sub>2</sub> enrichment on yield is dependent on the stage of growth at which it is applied. CO<sub>2</sub> enrichment at the early pod development stage did not increase soybean yield, while enrichment from the early pod development until maturity and from the emergence to maturity increased yields by 27 and 36% respectively (Ackerson *et al.*, 1984). A 20% increase in grain yield was recorded in wheat grown at elevated CO<sub>2</sub> under controlled environmental conditions (Mitchell *et al.*, 1996). In wheat Havelka *et al.*, (1984b) showed that

exposure to 1200 ppm had maximum effect during the period upto anthesis, while CO<sub>2</sub> enrichment increased yields of spring wheat most during tillering (Krenzer and Moss, 1975).-Kimball (1983) compiled more than 430 observations from 37 species and concluded that increasing CO<sub>2</sub> concentration to double the existing ambient levels will probably increase the yield of plant species. It is suggested that the additional carbohydrates formed with CO<sub>2</sub> enrichment are required during early flower and seed development and are not used during grain filling. Presumably the availability of energy and substrates determines how reproductive organs develop and the abundance of assimilate may allow young tillers and growing primordia to survive and produce more reproductive structures. Lack of proper understanding of factors determining the distribution of carbohydrates and nitrogenous compounds into sinks precludes prediction of the effects of CO<sub>2</sub> enrichment under a range of conditions.

### TISSUE COMPOSITION

The biochemical composition of plants undergo changes when grown at elevated CO<sub>2</sub> for a long period. Wheat accumulated double the amounts of sucrose and starch under elevated CO<sub>2</sub> (Havelka *et al.*, 1984b). Sweet potato had greater cellulose content in stems when grown under elevated CO<sub>2</sub>, but hemicellulose of leaves together with lignin in stems decreased (Bhattacharaya *et al.*, 1990). The tubers developed under elevated CO<sub>2</sub> contained less protein, total carotenoids and fibre and had a smaller percentage of nitrogen and protein. On the other hand, water content of tissues decreased at high CO<sub>2</sub> (Biswas and Hileman, 1985). Sweet potato grown in elevated CO<sub>2</sub> had more starch in storage tubers and leaves, but not in stems when compared with those grown under ambient CO<sub>2</sub> both under well watered and stress conditions (Bhattacharaya *et al.*, 1990).

There was no effect of water stress under CO<sub>2</sub> enrichment on the elemental composition of maize plants (Loomis and Lafitte, 1987). Similarly no CO<sub>2</sub> enrichment effect was observed on the moisture content, fibre, oil, proteins or fatty acid composition of maize and soybean seeds (Rogers *et al.*, 1983; 1986). Havelka *et al.*, (1984b) found no change in the nitrogen content per unit dry matter in leaves, stems or grain due to CO<sub>2</sub> enrichment in winter wheat. However, there are reports that the C:N ratio of cereals decreases, because of accumulation of carbohydrates and reduction in protein content. In experiments with

Scirpus, total soluble proteins, including Rubisco were decreased by 30-50% (Jacob *et al.*, 1994) which may have been the reason for the 15-20% reduction in the tissue N content (Curtis *et al.*, 1989). Similar results were obtained in rice by Rowland-Bamford *et al.*, 1991). Reduced tissue protein and N contents were also observed by workers (Thompson and Drake, 1994; Stitt, 1991; Hunt *et al.*, 1996). Changes in the tissue biochemical composition may affect the grain quality (Willimas *et al.*, 1995). Several studies have shown significant accumulation of carbohydrates in plants grown at elevated CO<sub>2</sub> (Rogers *et al.*, 1996a; 1996b; Stitt, 1991; Nie *et al.*, 1995a; Roden and Ball, 1996a).

Contradictory reports were made by Biswas and Hileman (1985) who noticed an increase in nitrogen content at elevated CO<sub>2</sub> conditions in cowpea. CO<sub>2</sub> had little effect on the water content of a range of plants including carrot, cotton, radish and soybean except under conditions which led to starch accumulation (Idso *et al.*, 1988). Changes in the tissue composition will alter the fodder quality of plants, nutritive value of grains and affect the foraging habits of herbivores, microbial decomposition, and therefore, the nutrient cycling through litter decomposition.

### SENESCENCE

Leaves grow faster and slightly earlier under elevated than normal ambient CO<sub>2</sub>. Therefore, leaves grown under elevated CO<sub>2</sub> conditions attain maturity faster than leaves grown at normal ambient CO<sub>2</sub>. There are reports showing high concentration of CO<sub>2</sub> decreasing the senescence process. Curtis *et al.*, (1989) reported slower rates of senescence and continued production of new shoots in *Scirpus olneyi* under elevated CO<sub>2</sub>. Mature Bartlett pea fruits when stored under a continuous flow of air with 10% CO<sub>2</sub> for 4 days showed reduced respiration and ethylene evolution rate and fruits remained greener and firmer than fruits stored in ambient air (Kerbel *et al.*, 1988). On the contrary, Bhattacharya *et al.*, (1985) found early leaf senescence in sweet potato grown at 675 ppm CO<sub>2</sub>. Senescence was not affected in soybean and wheat (Havelka *et al.*, 1984a; 1984b) and in White oak (Gunderson *et al.*, 1993) grown at elevated CO<sub>2</sub>. However, leaves of wheat plants grown in high CO<sub>2</sub> have been seen to senesce three to four days earlier (Sionet *et al.*, 1981). Recently Mc Connaughey *et al.* (1996) also found no change in senescence in two forest species.

High CO<sub>2</sub> probably has little direct effect on leaf senescence. The mechanism by which CO<sub>2</sub> might affect senescence is not clear. High levels (> 2000 ppm) inhibit the action of ethylene. Early senescence under elevated CO<sub>2</sub> may be correlated with the timing of other phenological events such as flowering (St. Omer and Horvath, 1983) or tuber maturation (Bhattacharya *et al.*, 1985).

### INTERACTION OF ATMOSPHERIC CO<sub>2</sub> WITH ENVIRONMENTAL FACTORS

Physiological or developmental responses of plants to increased CO<sub>2</sub> will depend strongly on other environmental factors. Global climate change is largely due to rising temperatures and uncertainty surrounding cloud formation and rainfall pattern. These will in turn alter the soil microbial activity which will alter the soil nutrient status and thus influence the response of plants to elevated CO<sub>2</sub>.

#### Temperature

Elevated CO<sub>2</sub> will increase photosynthesis while high temperature will increase photorespiration. For a given atmospheric CO<sub>2</sub>, an increase in temperature will increase the oxygenase activity of Rubisco relative to its carboxylase activity and thus the proportion of photosynthesis lost to photorespiration will be more at high temperature (Long 1991). Thus reduction in oxygenase by elevated CO<sub>2</sub> and hence increase in photosynthesis will have greater effect at high CO<sub>2</sub> rather than temperatures (Long 1991). Photosynthetic stimulation to elevated CO<sub>2</sub> has been more in summer than in winter (Lewis *et al.*, 1996). In the arctic tundra, doubling the atmospheric CO<sub>2</sub> only had a transient stimulation on photosynthesis (Oechel and Strain, 1985). But in a warm temperate climate, elevated CO<sub>2</sub> continued to stimulate photosynthesis and biomass production in *Scirpus olneyi* (Drake, 1992). Ziska *et al.*, (1990) showed that in this species, the relative effect of high CO<sub>2</sub> on photosynthesis was enhanced with a small temperature increase of 4 °C. This geographical variation in the results can be largely explained on the basis of the temperature differences. It is even predicted that boreal forests and tundra will emit CO<sub>2</sub>, but tropics will assimilate carbon in a future world rich in CO<sub>2</sub> (Wang and Polgalse, 1995). However, Udayakumar and Prasad (1993) have argued that the beneficial effects of elevated CO<sub>2</sub> on photosynthesis will be nullified at high temperatures. In this context it may be noted that Sionet *et al.*, (1987) reported increased leaf



photosynthetic rates of soybean in response to elevated CO<sub>2</sub> more at 22/16 °C than at 26/20 °C.

Increased temperature enhanced the growth at elevated CO<sub>2</sub> in various species such as the tropical aquatic plant *Eichornia crassipes* (Idso *et al.*, 1987), cassava (Imai *et al.*, 1984) and soybean (Sionit *et al.*, 1987). Temperature and CO<sub>2</sub> had opposite effects on growth of determinate crops such as wheat (Mitchell *et al.*, 1996).

Elevated CO<sub>2</sub> concentrations can affect plant growth and development at subtropical temperatures. Orange trees grown at 800 ppm CO<sub>2</sub> before anthesis, showed an increase in fruit set (Downton *et al.*, 1987). Growth at elevated CO<sub>2</sub> reduced chilling inhibition of carbohydrate transport in two species of temperate C4 grasses (Potvin, 1985). At sub-optimal temperature, okra was found to have higher survival, growth and reproduction at elevated CO<sub>2</sub> than normal ambient CO<sub>2</sub> levels (Sionit *et al.*, 1981). Rogers *et al.*, (1984) noticed a marginal decrease in seed yield of soybean at 660 ppm CO<sub>2</sub> with temperature range of 23 to 33°C, however seed yield increased with temperature at 330 ppm CO<sub>2</sub>. Ryle *et al.*, (1992) found that combined effect of CO<sub>2</sub> and temperature reduced the root to shoot ratio of white clover.

### Light

Higher photosynthetic rates under elevated CO<sub>2</sub> are highly dependent on light. Light saturation is usually increased under elevated CO<sub>2</sub> compared with ambient CO<sub>2</sub> (Valle *et al.*, 1985; Wong and Dunin, 1987; Campbell, 1988). Soybean plants grown at high CO<sub>2</sub> exhibited better capability to utilise radiation effectively throughout all light levels than plants grown at low CO<sub>2</sub> (Valle *et al.*, 1985). Light compensation point was reduced from 35  $\mu$ moles photons m<sup>-2</sup> s<sup>-1</sup> at 300 ppm CO<sub>2</sub> to 27  $\mu$ moles photons m<sup>-2</sup> s<sup>-1</sup> at 660 ppm CO<sub>2</sub>. Sharon and Mitchell (1988) found lettuce leaf growth to respond most to a combination of high light intensities and CO<sub>2</sub> enrichment. Plants exposed to elevated CO<sub>2</sub> can utilize low light better than those grown at normal ambient CO<sub>2</sub> (Long and Drake, 1991). The apparent quantum yield for photosynthesis increased by 93% in *Quercus* plants grown at elevated CO<sub>2</sub> (Scarasciia-Mugnozza *et al.*, 1996). In tropical forests where photon flux and water availability are high (Chazdon and Fetcher, 1984; Becker *et al.*, 1988) plants may be able to express their full physiological potential when exposed to elevated concentration of CO<sub>2</sub>.

### Nutrients

Plant responses to elevated CO<sub>2</sub> can vary depending on the nutrient status of the soil. One opinion is that plants can perform better under elevated CO<sub>2</sub> levels even if the soil nutrient status is relatively poor, while another view is that continued response of plants to elevated CO<sub>2</sub> needs higher nutrients as biomass production increases drastically at elevated CO<sub>2</sub> (Wong *et al.*, 1992).-

Wong *et al.* (1979) have shown that the response to N fertilizer under elevated CO<sub>2</sub> is species specific. In their experiment with four different Eucalyptus species they have shown a positive interaction with N. Number of leaves per plant increased substantially with high CO<sub>2</sub> when combined with high N. Nevertheless, the nature of allocation of nutrients to different plant parts depends on the nutrient status. Nutrient rich condition will call for less root to service a given shoot mass, with a consequent increase in shoot:root ratio (Wilson, 1988; Wong, *et al.*, 1992). Shoot biomass increased by 30% at elevated CO<sub>2</sub> irrespective of N supply, but growth effects closely matched changes in sink development mediated through N supply (Rogers *et al.*, 1996a; 1996b). In rice photosynthesis and growth response to elevated CO<sub>2</sub> were highly dependent on N supply and was mediated through sink development (Ziska *et al.*, 1996). Elevated CO<sub>2</sub> increased photosynthesis in K deficient needles of Norway Spruce (Barnes *et al.*, 1995).

### Moisture Stress

Under elevated CO<sub>2</sub> conditions stomatal conductance decreases, and therefore, water loss through stomata decreases.-It is clear from several studies on temperate species that increased CO<sub>2</sub> can substantially reduce water loss primarily through stomatal closure (Rogers *et al.*, 1983; 1984). Tolly and Strain (1985) found that for the temperate tree Liquidamber styraciflua and Pinus taeda, elevated CO<sub>2</sub> delayed the onset of water stress effects and allowed the plants to maintain high photosynthetic rates for a longer period of time. The leaf water potentials at which photosynthesis was inhibited in ambient air grown plants was not affected by the CO<sub>2</sub> treatment, indicating that the effect was most likely due to CO<sub>2</sub> effects on conductance, and therefore, on transpiration. In a similar study, Gifford (1979) also noted that water stress enhanced the positive effect of elevated CO<sub>2</sub> on plant biomass.

Elevated CO<sub>2</sub> increased the leaf water potential in Eucalyptus during drought and alleviated the effects of drought (Roden and Ball, 1996a; Scarascia-Mugnozza *et al.*, 1996). Growth at elevated CO<sub>2</sub> reduced the oxidative stress (Schwanz *et al.*, 1996) and even improved the PSII activity which is very sensitive to drought in plants exposed to elevated CO<sub>2</sub> (Scarascia-Mugnozza *et al.*, 1996). C<sub>4</sub> photosynthesis responded positively to elevated CO<sub>2</sub> during drought but not under well watered conditions (Samarakoon *et al.*, 1996a). Elevated CO<sub>2</sub> increased the WUE more in drying than well watered cotton plants (Samarakoon *et al.*, 1996b). Irrigation requirements for wheat have been predicted to decline with rising CO<sub>2</sub> concentration (Senock *et al.*, 1996).

## ELEVATED CO<sub>2</sub> AND ECOSYSTEM PROCESSES

### Biomass

The higher rates of photosynthesis and lower rates of respiration observed in single leaf measurements were translated into field level measurable effects. Measurements of ecosystem carbon uptake rate and respiration showed a considerable increase and decrease, respectively, in plants grown at elevated CO<sub>2</sub> in the field for several years (Long and Drake, 1992). This led to increased biomass produced per unit land area as discussed earlier. Reduced stomatal conductance and transpiration led to better water use efficiency in term of the amount of water consumed to produce a given amount of biomass on a land area basis compared to plants grown at normal ambient CO<sub>2</sub>.

Continuous growth at elevated CO<sub>2</sub> in the field for several years led to sustained stimulation in canopy photosynthesis and increased net primary production of both shoot and root systems in plant communities dominated by C<sub>3</sub> plants which resulted in accumulation of carbon in the soil (Drake 1992). Increased soil carbon might have increased the soil microbial activity leading to increased production of methane in plots exposed to elevated CO<sub>2</sub> for several years (Dacey *et al.*, 1994). Increased soil microbial biomass carbon due to elevated CO<sub>2</sub> has been reported by Schenk *et al.*, (1995).

### Insects and Fungi

Increased availability of photosynthates and decrease in the tissue protein content leads to changes in the C:N ratio which is an important

factor determining the forage quality and degradability of the tissue during microbial decomposition. A reduction in the nutritive value of the plants from the reduction in their protein contents (and thus, of tissue N) may affect the foraging habits of herbivores and fungi in a future world rich with CO<sub>2</sub> (Thompson and Drake, 1994). Insect larvae have been shown to increase the tissue consumption to maintain their total N uptake when fed with N poor tissue (Scriber and Slansky, 1981). However, their growth is often markedly reduced (Price, 1984). Many types of larvae exhibited increased consumption of plant tissues grown at elevated CO<sub>2</sub> (Lincoln *et al.*, 1986; 1989; Fajer *et al.*, 1989) and showed early instar development (Fajer *et al.*, 1989). Thompson and Drake (1994) found that fewer plants grown in elevated CO<sub>2</sub> were attacked by insects and even in those plants infested by the insects less tissue was eaten by the larvae and that the tissue N content correlated positively with the insect infestation. Due to poor nutrient status, the female larvae of beech weevil ate in a compensatory manner, but the male larvae were not affected (Docherty *et al.*, 1996).

Reduced tissue N content has been cited as the reason for reduced fungal infection of wheat grown at elevated CO<sub>2</sub> (Thompson *et al.*, 1993). However in the same study they found that wheat plants grown at elevated CO<sub>2</sub> and subjected to water stress did not reduce their tissue N content but increased the tissue water content and that caused increased fungal attack. Thompson and Drake (1994) reported a significant reduction in the severity of fungal infestation of plants grown at elevated CO<sub>2</sub> only when the tissue N content was less. In some C4 plants where the tissue N did not decrease, better relative water content of the tissue led to increased fungal attack (Thompson and Drake, 1994). Thus the degree of fungal infestation may depend on the tissue composition and water content. Also, changes in the microhabitat, particularly relative humidity resulting from changed stomatal conductance and transpiration may also play a role in fungal and insect population dynamics and hence on the extent of damage that plants may suffer from insects and fungus in a future world with elevated CO<sub>2</sub>. It is possible that the microbial population also may show a tissue sensitive response which may affect the rate of litter decomposition and nutrient cycling in forests and agricultural ecosystems.

### INFLUENCE OF ELEVATED CO<sub>2</sub> ON PLANT-PLANT INTERACTION

In order to know the plant community response to elevated CO<sub>2</sub>, it is necessary to understand how interactions among neighboring plants in a plant community would alter the growth and reproduction of individual species.

In a study where a community of plant species comprising of both C<sub>3</sub> and C<sub>4</sub> species was exposed to elevated CO<sub>2</sub> concentration, the C<sub>3</sub> species showed higher growth rates than C<sub>4</sub> species (Carter and Paterson, 1983). Such a species response was later shown to be mainly due to the differences in their photosynthetic response to elevated CO<sub>2</sub> (Paterson *et al.*, 1984). C<sub>3</sub> plants being more sensitive to changes in ambient CO<sub>2</sub> concentration may out grow C<sub>4</sub> species in a community (Drake, 1992). Due to differences in the species response to elevated CO<sub>2</sub>, community structure may change in the native ecosystems (Leadley and Stocklin, 1996) such as grass lands and forests.-

Changes in growth rate can create a competition among the various species of a community for natural resources like light, moisture, nutrients, pollinators and microclimatic conditions like humidity, temperature and wind (Bazzaz and carlson, 1984; Zangler and Bazzaz, 1984). Therefore, apart from photosynthetic responses of C<sub>3</sub> and C<sub>4</sub> species, the competition for natural resources will bring in plant-plant interaction (Bazaz *et al.*, 1992) and may lead to composition changes in the community.

Another possible way through which elevated CO<sub>2</sub> concentration might affect the plant-plant interaction is through the changes that it brings about in the tissue composition. We have seen that CO<sub>2</sub> affects C:N ratio significantly in addition to other tissue components. This can lead to differential preference of insects and pests and there by some species become more susceptible in multiple species stands. Pollinators that frequent a species might also change based on the nectar secreting ability the nutritive value of which may be again altered due to growth in elevated CO<sub>2</sub>. This can affect the reproductive fitness of certain species. As a cumulative effect, after a few generations this can lead to changes in the species composition of a community.-When such a differential growth response occurs, certain species may become locally extinct after several generations following the Darwinian laws of natural selection.



## CONCLUSIONS

This review reveals how molecular, physiological and environmental factors regulate carbon metabolism in vegetation grown at elevated concentrations of CO<sub>2</sub> and how that will have a strong impact on global carbon cycle. Annually, global photosynthesis and respiration cycles more than 100 GT of CO<sub>2</sub> between the atmosphere and the pools of carbon in terrestrial ecosystems (Gifford, 1994; Goudriaan, 1993). Analysis of the major sources and sinks for carbon in the biosphere suggests that a net 0.4-2.4 Gt/year uptake into terrestrial ecosystems may be the result of CO<sub>2</sub> fertilization caused by increasing atmospheric CO<sub>2</sub> since 1958 (Gifford, 1994; Goudrian, 1993). The mechanisms regulating carbon flow in ecosystems in response to rising CO<sub>2</sub> and climate change will determine the future levels of atmospheric CO<sub>2</sub>.

Majority of the CO<sub>2</sub> experiments have been conducted in controlled environments, and the lack of agreement between various studies is mainly because of the varying conditions under which the experiments were conducted. Different types of systems such as closed structures, open top chambers, free air CO<sub>2</sub> enrichment (FACE) system etc are used in CO<sub>2</sub> experiments. In majority of the CO<sub>2</sub> experiments potted plants are used.-A major inadequacy of pot experiments is the small volume of soil available to roots which necessitates frequent watering and application of nutrient. The small soil volume in pots may restrict the growth for many reasons which cannot be over come by application of water and nutrients in more split doses (Lawlor & Mitchell, 1991; Arp, 1991; Radin *et al.*, 1987; Arp and Drake, 1991; Campbell *et al.*, 1988). One must take all these factors in to consideration while comparing or making any conclusions from the different CO<sub>2</sub> studies conducted by different researchers under different growth conditions.

There are a number of models developed to predict future growth and yield of plants in the agricultural and natural ecosystems and the changes in ecosystem and climate processes that are associated with the increasing atmospheric CO<sub>2</sub> concentrations. Predictions using models that are based on valid scientific data, both meteorological and biological will be useful to develop guidelines for future planning.

Productivity of vegetation is an outcome of highly complicated interactions of many environmental, soil, water, nutrient and plant factors. Interaction of environmental factors plays a key role in regulating the physiological processes of plants to elevated atmospheric

CO<sub>2</sub> concentration. The compositional changes in the tissue due to elevated CO<sub>2</sub> concentration can bring in changes in the plant community making some plants more vulnerable and some less to insect/pest damage. In addition to this, the varying species response to elevated CO<sub>2</sub> concentration in terms of growth may cause changes in the composition of the plant community in a few generations of a species growth at elevated concentrations of atmospheric CO<sub>2</sub>.

There is a general expectation that rising CO<sub>2</sub> and increasing global temperature will be accompanied by increased production of plant biomass. This is largely based on the well known kinetic responses of photosynthesis, growth and water use efficiency to increased atmospheric CO<sub>2</sub> concentration. Changes in respiration rates can also contribute significantly in the overall carbon budget of a plant. Because more than 50% of the carbon assimilated through photosynthesis is lost in respiration in the life time of a plant. The physiological and biochemical processes of vegetations have a pivotal role in global carbon cycle. The ability of plants to put on more biomass under elevated atmospheric CO<sub>2</sub> concentration can be expected to have a positive impact on enhancing growth rates of crop plants and agricultural productivity.

## REFERENCES

1. Ackerson, R.C., Havelka, U.D. and Boyle, M.G. 1984. CO<sub>2</sub> enrichment effects on soybean physiology. II effects of stage specific CO<sub>2</sub> exposure. *Crop Sci.* 24, 1150-1154.
2. Allen, L. H. Jr., 1990, Plant responses to rising carbon dioxide and potential interactions with air pollutants. *J. Environ. Qual.* 19: 15-34.
3. Amthor, J.S. 1991. Respiration in a future, higher CO<sub>2</sub> world. *Plant, cell and environment*, 14, 13-20.
4. Arp, W.J. 1991. Effects of source-sink relations on- photosynthetic acclimation to elevated CO<sub>2</sub>. *Plant, cell and environ.* 14, 869-875.
5. Arp, W.J. and Drake. B.G. 1991. Increased photosynthetic capacity of *Scripus olneyi* after 4 years of exposure to elevated CO<sub>2</sub>. *Plant, cell and Environ.* 14, 1004- 1008.
6. Azcon-Bieto J., Gonzalez-Meler M.A., Doherty W. and Drake B.G. 1994. Acclimation of respiratory O<sub>2</sub> uptake in green tissues of field-grown native species after long-term exposure to elevated atmospheric CO<sub>2</sub>. *Plant Physiol.*
7. Azcon-bieto, J. and Osmand, B.C. 1983. Relationship betwnn photosynthesis and respiration. *Plant Physiol.* 71, 574-581.

8. Baker, J. T., Allen, L. H., Boote, K. J., Jones, P. and Jones, J. W., 1989, Response of soybean to air temperature and carbon dioxide concentration. *Crop Sci.*, 29: 98-105.
9. Baker, J. T., Allen, L. H., Jr. and Boot, K. J., 1990, Growth and responses of rice to CO<sub>2</sub> concentrations. *J. Agric. Sci.*, 115: 313-320.
10. Barnes, J.D., Pfirrmann, T., Steiner, K., Lutz, C., Busch, U., Kuchenhoff, H., Payer, H. D., 1995. Effects of elevated CO<sub>2</sub>, elevated O<sub>3</sub> and potassium deficiency on Norway spruce (*Picea abies* L. Karst) : Seasonal changes in Photosynthesis and non-structural carbohydrate content. *Plant. Cell Environ.* 18, 1345-1357.
11. Bazzaz, F.A., and Carlson, R.W. 1984. The response of plants to elevated CO<sub>2</sub>-I Competition among an assemblage of annuals at two levels of soil moisture. *Oecologia*, 62, 196-198.
12. Bazzaz, F.A., and Kelly, D.M. McCaughay, 1992. Plant - plant interactions in elevated CO<sub>2</sub> environments. *Aust. J. Bot.*, 40, 547-563.
13. Becker, P.F., Rabenold, P.E. Idol, J.A. and Smith. A.P. 1988. Water potential gradients for gaps and slopes in a panamerican tropical moist forest's dry season. *Journal of tropical Ecology*, 4, 173-184.
14. Besford, T. 1993. Photosynthetic acclimation in tomato plants grown in high CO<sub>2</sub>. In CO<sub>2</sub> and Biosphere. Eds. J Rozema, H. Lambers, S.C. Van de Gein & M.L. Cambridge. Kluwer Academic Publishers, london. pp. 441-448.
15. Bhattacharya, M.C., Biswas, P.R., Bhattacharya, S., Sionit, N. and Strain, B.R., 1985, Growth and Yield response of sweet potato to atmospheric CO<sub>2</sub> enrichment. *Crop. Sci.*, 25(6):---- 975-981.
16. Bhattacharya, M.C., Hileman, D.R., Gosh. P.P., Musser, R.L., Bhattacharaya, S. and Biswas, P.K. 1990. Interaction of enriched CO<sub>2</sub> and water stress on the physiology and biomass production in sweet potato grown in open top chambers. *Plant, cell and environ.*, 13, 933-940.
17. Biswas, P.K. and Hileman, D.R. 1985. Response of vegetation to carbon dioxide: field studies of sweet potatoes and coepeas in response to elevated carbon dioxide. Report 022, US Dept. of energy, carbon dioxide research Division, Office of energy research, Washington, DC.
18. Bowes, G. 1993. Facing the inevitable: Plants and increasing atmospheric CO<sub>2</sub>. *Annl. Rev. Plant Physiol. & Plant mol. biol.* 44:309-332.
19. Bunce, J. A., 1990, -Short- and long-term inhibition of respiratory carbon dioxide efflux by elevated carbon dioxide. *Annal. Bot.*, 65: 637-642.
20. Calvert, A and Slack, G., 1975, Effects of CO<sub>2</sub> enrichment on growth, development and yield of green house tomatoes. I Response to controlled conditions. *J. Hort. Sci.*, 50: 61-71.

21. Campbell, W. J., Allen, L. H. and Bowes, G., 1988, Effects of CO<sub>2</sub> concentration on Rubisco activity, amount and Photosynthesis in soybean leaves. *Plant Physiol.*, 88: 110-1316. f CO<sub>2</sub>
22. Carter, D.R. and Patterson, K.M., 1983. Effects of a CO<sub>2</sub> enriched atmosphere on the growth and competitive interactions of a C3 & C4 grass. *Oecologia (Berlin)*. 58, 188-193.
23. Cave, G., Tolley, L. C. and Strain, B. R., 1981, Effect of CO<sub>2</sub> enrichment on chlorophyll----content, starch content and starch grain structure in *Trifolium* leaves. *Plant Physiol.*, 88: 1310-1316.
24. Chaudhuri, U.K., Kirkham, M.B. and Kanemaru, E.T. 1990. Root growth of winter wheat under elevated carbon dioxide and drought. *Crop Science*. 30,853-857.
25. Chazdon, R.L. and Fetcher, N. 1984. Photosynthetic light environments in a low land tropical forest in Costa Rica. *J.Ecol.* 72, 553-564.
26. Clough, J.M., Peet, M.M. and Kramer. P.J. 1981. Effect of high atmospheric CO<sub>2</sub> and sink on size on rates of photosynthesis of a soybean cultivar. *Plant Physiol.* 67, 1007-1010.
27. Cure, J. D. and Acock, B., 1986,-Crop responses to carbon dioxide doubling : a literature survey. *Agric. Forest Meteorol.*, 38: 127-145.
28. Cure, J.D., Rufty, T.W.Jr. and Israel, D.W. 1989. Alterations in soybean leaf development and photosynthesis in a CO<sub>2</sub>-enriched atmosphere, *Bot. Gazz.*, 150, 337-345.
29. Curtis, P. S., Drake, B. G., Leadley, P. W., Arp, W. J. and Whigham, D. F., 1989, Growth and senescence in plant communities exposed to elevated CO<sub>2</sub> concentrations on an estuarine marsh. *Oecologia*, 78: 20-26.
30. Curtis, P.S. and Teeri, J.A. 1992. Seasonal responses of leaf gas exchange to elevated CO<sub>2</sub> in *Populus grandidentata*. *Cann. J.Forest Res.* 22, 1320-1325.
31. Day, F.P., Weber, E.P, Hinkle, C.R. and Drake, B.G. 1996. Effects of elevated atmospheric CO<sub>2</sub> on fine root length and distribution in an oak-palmetto scrub ecosystem in central Florida. *Global Change Biol.* 2:143-148.
32. Dacey, J.C., Drake, B.G. and M. Klug. 1994. Elevated CO<sub>2</sub> increases methane emission from a Chesapeake Bay wet land. *Nature*, 370: 47-49.
33. Del Castillo, D. Acock, B, Reddy, V.R. and Acock, M.C. 1989. Elongation and branching o roots on soybean plants in a carbon dioxide enriched aerial environment. *Agron. J.* 81, 692-695.
34. DeLucia, E.H., Sasek, T.W. and Strain, B.R., 1985, Photosynthesis inhibition after long term exposure to elevated levels of atmospheric carbon dioxide. *Photosynthesis Research.*, 7: 175-184.

35. Devakumar A.S. 1994. Response of tree species to elevated CO<sub>2</sub> : An approach to enhance early sapling growth rate. Ph.D. thesis, Department of Crop Physiology, University Of Agricultural Sciences, Bangalore.
36. Devakumar A.S., Jacob. J. and Sethuraj, M.R. 1996. A novel approach to obtain increased growth in nursery seedlings of *Hevea brasiliensis* using CO<sub>2</sub> fertilization. Abstract of International Rubber Research and Development Board Symposium on Agronomy and Farming systems, Sri Lanka, pp.2.
37. Diaz, S., Grime, J.P., Harris, J. and McPherson, E. 1994. Evidence of feedback mechanism limiting plant response to elevated CO<sub>2</sub>. *Nature* 364-317.
38. Docherty M., Hurst D.K., Holopainen J.K., Whittaker J.B., Lea P.J. and Watt A.D., 1996. Carbon dioxide induced changes in beech foliage cause female beech weevil larvae to feed in a compensatory manner. *Global Change Biology*, 2, 335-341.
39. Downton, W.J.S, Grant, W.J.R and Loveys, B.R., 1987, CO<sub>2</sub> enrichment increases yield of Valencia orange. *Aust. J. Plant Physiol*, 14: 493-501.
40. Drake B.G. 1992. A field study on the effects of elevated CO<sub>2</sub> on ecosystem processes in a Chesapeake bay wet land. *Aust. J. Biol.* 40:579-95.
41. Fajer, E.D., Bowers. M.D., and Bazzaz, F.A. 1989. The effects of enriched carbon dioxide atmospheres on plant-insect herbivore interactions. *Science*, 243, 1198-1200.
42. Farrar, J.F., and Williams, M.L., 1991. The effects of increasing atmospheric carbon dioxide and temperature on carbon partitioning, source sink relations and respiration. *Plant Cell Environ.* 14, 819-830.
43. Farquhar, G.D., von Caemmerer, S. and Berry, J.A. 1980. A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* 149: 78-90.
44. Field, C.B., Jackson, R.B. and Mooney H.A. 1995. Stomatal responses to increased CO<sub>2</sub> : implications from the plant to the global scale. *Plant Cell. Environ.* 18, 1214-1225.
45. Gifford, R.M. 1994. The global carbon cycle: A view point on the missing sink. *Asut. J. Plant Physiol.* 21, 1-15.
46. Gifford, R.M., 1979, Growth and yield of CO<sub>2</sub> enriched wheat under water limited conditions. *Aust. J. plant Physiol.*, 6: 367-368.
47. Gonzalez M.A., Carbo M.R., Siedow J.M., and Drake B.G. 1996. Direct inhibition of plant mitochondrial respiration by elevated CO<sub>2</sub>. 112, 1349-1355.
48. Gunderson C.A., Norby R.L. and Wullschlegel S.D. 1993. Foliar gas exchange responses of two deciduous hardwood during 3 years of growth in elevated CO<sub>2</sub> : no loss of photosynthetic enhancement. *Plant, Cell and Environ.* 16: 770-778.



49. Gourdiaan, J. 1993. In: S.C. van de geijn, J. Gourdiaan & F.Berendse (eds) Climate Change: crops and terrestrial ecosystems. CABO-DLO, Wageningen.
50. Gourdian, J and de Ruiter, H.E., 1983. Plant growth in response to CO<sub>2</sub> enrichment at two levels of nitrogen and phosphorus supply. I. Dry matter, leaf area and development. *Neth. J. Agric. Sci.*, 31, 157-169.
51. Gustafson, S.W. 1984. Effects of CO<sub>2</sub> enrichment during flowering and pod filling on net photosynthesis; dry matter accumulation and yield of beans, *Phaseolus vulgaris*. L. Dissertation abstracts International. 44 : 2954.
52. Havelka, U. D., Ackerson, R. C., Boyle, M. G. and Wittenbach, L. A., 1984a, CO<sub>2</sub> enrichment effect on soybean physiology I. Effects of long-term exposure. *Crop Sci.*, 24: 1146-1168.
53. Havelka, U. D., Wittenbah, V. A. and Boyle, M. G., 1984b, CO<sub>2</sub>-enrichment effects on wheat yield and physiology. *Crop Sci.*, 24: 1163-1168.
54. Heij, G., Uffelen, J.A.M and Van., 1984, Effects of CO<sub>2</sub> concentration on growth of glass house cucumber *Acata Hort.*, 162: 29-36.
55. Hendrix, D.L. Mauney, J.R., Kimball, B.A., Lewin, K.F., Nagy, J. and Hendrey, G.R., 1994. Influence of elevated CO<sub>2</sub> and mild water stress on non-structured carbohydrates in field grown cotton tissues. *Agric. & Forest Meteorol.* 70, 153-162.
56. Hrubec, T. C., Robinson, J. M. and Donaldson, R. P., 1985, Effect of CO<sub>2</sub> enrichment and carbohydrate content on the dark respiration of soybeans. *Plant Physiol.*, 79: 684-689.
57. Huber, S.C., Rogers, H.H. and Mowry, F.L. 1984. Effects of water stress on photosynthesis and carbon partitioning in soybean plants grown in the field at different CO<sub>2</sub> levels. *Plant Physiol.* 76, 244-249.
58. Hunt H.W., Elliot E.T., Detling J.K., Morgan J.A. and Chen D.X., 1996. Response to elevated CO<sub>2</sub> and temperature under different water regimes, *Global Change Biology.* 2, 35-47.
59. Idso, S. B., Kimball, B. A., Anderson, M. G. and Mauney, J. R., 1987,- Effects of atmospheric CO<sub>2</sub> enrichment on plant growth: the interactive role of air temperature. *Agric. Eosyst. Environ.*, 20: 1-10.
60. Idso, S. B., Kimball, B. A., and Mauny, J. R. 1988. Atmospheric CO<sub>2</sub> enrichment and plant dry matter content. *Agric. forest meteorol.*, 43, 171-181.
61. Idso, I.B. and Kimbell, B.A., 1991, Effects of two and a half years of atmospheric CO<sub>2</sub> enrichment on the root density distribution of tree-year-old sour orange trees. *Agric. and Forest Meteorol.*, 55: 345-349.
62. Idso, S.B. and Kimball, B.A. 1992. Seasonal fine root biomass development of sour orange trees grown in atmospheres of ambient and elevated CO<sub>2</sub> concentrations. *Plant Cell Environ.* 15, 337-341.

63. Idos, S.B. and Kimball, B.A. 1994. Effects of atmospheric CO<sub>2</sub> enrichment on regrowth of sour orange trees (*Citrus aurantium*) after cooiecing. Am. J. Bot. 81(7): 843-846.
64. Imai, K., Coleman, D. F. and Yagagisava, T., 1984, Elevated atmospheric partial pressure of CO<sub>2</sub> and dry matter production of cassava (*Manihot esculenta*, Crantz.). Japanese J. Crop Sci., 53: 479-485.
- ✓65. Jacob, J. Grieter, C., and Drake, B.G. 1995. Acclimation of photosynthesis in relation to Rubisco and nonstructural carbohydrate contents and *in situ* carboxylase activity in *Scripus olneyi* grown at elevated CO<sub>2</sub> in the field. Plant Cell Environ. 18, 875-884.
66. John, T.A., and Lloyd, J. 1992. Sources and sinks of atmospheric CO<sub>2</sub>. Aust. J. Bot. 40, 407-418.
67. Jones, P.G., Lloyd, J.C. and Raines, C.A. 1996. Glucose feeding of intact wheat plants represses the expression of a number of calvin cycle genes. Plant. Cell. Environ., 19, 231-236.
68. Jones, P. Jones, J. W. and Allen, L. H. Jr., 1985, CO<sub>2</sub> effects on photosynthesis and transpiration during vegetative growth in soybeans. In : Proceedings of soil and Crop---Science Society of Florida, 44: 129-134.
69. Jordan, D.B. and Ogren, W.L. 1984. The CO<sub>2</sub> specificity of ribulose 1,5-bisphosphate carboxylase/oxygenase. Planta. 161, 308-313.
70. Kerbel, E. L., Kader, A. A. and Romani, R. J., 1988, Effects of elevated CO<sub>2</sub> concentrations on glycolysis in intact 'Bartlett' pear fruits. Plant Physiol., 86: 1205-1209.
71. Kimball, B. A. 1983. Carbon dioxide and agricultural yield: an assemblage and analysis of 430 prior observations. Agronomy journal, 75, 779-788.
72. Kimball, B.A. and Idso, S.B. 1983. Increasing atmospheric CO<sub>2</sub>: Effects on crop yield, water---use and climate. Agril. Water management. 7, 63-74.
73. Krenzer, E.G. and Moss, D.N., 1975. Carbon dioxide enrichment effects upon yield and yield components. Crop Sci., 15: 71-74.
74. Kriedmann, P.E., Sward, R.J. and Downtown, W.J.S. 1976. Vine response to CO<sub>2</sub> enrichment during heat therapy. Aust. J. Plant Physiol. 3, 605- 618.
75. Kroner, C. H., Diemer M. 1994. Evidence that plants from high altitudes retain their photosynthetic efficiency under elevated CO<sub>2</sub>. Functional Ecology 8, 58-68.
76. Ku, S.B. and Edwards, G.E., 1977. Oxygen inhibition of photosynthesis II. Kinetic characteristics as affected by temperature. Plant Physiol., 59: 991-999.
77. Lawlor, D.W. and Mitchel, A.R.C., 1990. The effects of increasing CO<sub>2</sub> on crop photosynthesis and productivity: A review of field studies. Plant Cell Environ., 14: 807-818.

78. Lawlor, D.W. and R.A.C. Mitchell. 1991. The effects of increasing CO<sub>2</sub> on crop photosynthesis and productivity: a review of field studies. *Plant. Cell and Environ.* 14, 807-818.
79. Leadly P.W. and Stocklin J., 1996. Effects of elevated CO<sub>2</sub> on moist calcareous grasslands : community, species and genotype level responses. *Global Change Biology*, 2,389-397.
80. Leadly, P.W. and Reynolds, J.F., 1988. Effects of elevated CO<sub>2</sub> on estimation of leaf area and leaf by weight of soybean. *Amer. J. Bot.*, 75: 1771-1774.
81. Leith, Y.H., Reynolds, J.F. and Rogers, H.H., 1986. Estimation of leaf area of soybeans grown under elevated CO<sub>2</sub> levels. *Field Crop Res.*, 13: 193-203.
82. Lewis J.D., Tissue D.T. and Strain B.R. 1996. Seasonal response of photosynthesis to elevated CO<sub>2</sub> in loblolly pine (*Pinus taeda* L.) over two growing seasons. *Global Change Biology*, 2,103-114.
83. Lincoln, D.E., Couvet, D., and Sionit, N. 1986. Response of an insect herbivore to host plants grown in carbon dioxide enriched atmospheres. *Oecologia*. 69, 556-560.
84. Long, S.P. 1991. Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO<sub>2</sub> concentration : Has its importance been underestimated? *Plant. Cell Environ.* 14,729-739.
85. Long, S.P. and Drake, B.G. 1991. Effect of the long term elevation of CO<sub>2</sub> concentration in the field on the quantum yield of photosynthesis of the C<sub>3</sub> sedge *Scripus olneyi*. *Plant Physiology* 96: 221-226.
86. Long S.P. and Drake, B.G. 1992. Photosynthetic CO<sub>2</sub> assimilation and rising atmospheric CO<sub>2</sub> concentrations. In *Crop Photosynthesis: Spatial and temporal Determinants* (eds N.R. Baker & H.Thomas), pp 69-103. Elsevier Science Publishers.
87. Loomis, R.S., and Lafitte, H.R. 1987. The carbon economy of a maize crop exposed to elevated CO<sub>2</sub> concentration and water stress as determined from elemental analysis. *Field Crop Res.* 17, 63-74.
88. Majeau N. and Coleman J.R. 1996. Effect of CO<sub>2</sub> concentration on carbonic anhydrase/ Oxygenase expression in pea. 112, 569-574.
89. Marc, J. and Gifford, R.M., 1984. Floral initiation in wheat, sunflower and sorghum under carbon dioxide enrichment. *Can. J. Bot.*, 62: 9-14.
90. Mc Connaughy, K.D.M., Bassow, S.L., Bernlson and Bazzaz, F.A., 1996. Leaf senescence and decline of end of season gas exchange in five temperate deciduous tree species grown in elevated CO<sub>2</sub> concentration. *Global Change Biology*, 2, 25-34.

91. Micallef, B.J., Vanderveer, P.J. and Sharkey, T.D., 1996. Responses to elevated CO<sub>2</sub> of flaveria linearis plants having a reduced activity of cytosolic fructose-1,6-bisphosphatase. *Plant. Cell. Environ.* 19, 10-16.
92. Miglietta, F.A., Raschi, R., Resti, and Badiani, M., 1993. Growth and onto-morphogenesis of soybean in an open, naturally, CO<sub>2</sub> environment. *Plant Cell Environ.*, 16: 909-918.
93. Mitchell, R.A.C., Gibbard, C.L., Mitchell, V.J. and Lawlor, D.W., 1996. Effects of shading in different developmental phases on biomass and grain yield of winter wheat at ambient and elevated CO<sub>2</sub>. *Plant. Cell. Environ.* 19, 615-621.
94. Morison, J.L., 1985, Sensitivity of stomata and water use efficiency to high CO<sub>2</sub>. *Plant, Cell and Environ.* 8: 789-796.
95. Neals, T.F. and Nicholls, A.O. 1978. Growth response of young wheat plants to a range of ambient CO<sub>2</sub> levels. *Aust. J. Plant. Physiol.* 5, 45-59.
96. Neftel, A., Moor, E., Oeschger, H and Stuffer, B. 1985. Evidence from polar ice cores for the increase in atmospheric CO<sub>2</sub> in the two contrives. *Nature*, 315: 45-47.
97. Nie, G., Handrix, D.L. Webber, A.N., Kimball, B.A. and Long, S.P. 1995a. Increased accumulation of carbohydrates and decreased photosynthetic Gene transcripts levels in wheat grown at an elevated CO<sub>2</sub> concentration in the field. *Plant Physiol.*, 108: 975-983.
98. Nie, G.Y., Long, S.P., Gracia, R.L., Kimball, B.A., Lamorte, R.L., Pinter Jr, P.J., Wall, G.W., and Webber, A.N., 1995b. Effects of free air CO<sub>2</sub> enrichment on the development of the photosynthetic apparatus in wheat, as indicated by changes in leaf proteins. *Plant. Cell. Environ.* 18. 855-864.
99. Nijs, I., Impens and Behaeghe, T., 1989, Effect of different CO<sub>2</sub> environments on the photosynthesis- yield relationships and the carbon and water balance of a white clover sward. *J. Expt. Bot.*, 40: 353-359.
100. Nobel, P.S. Israel, A.A., and Wang, N., 1996. Growth, CO<sub>2</sub> uptake, and responses of the carboxylating enzymes to inorganic carbon in two highly productive CAM species at current and doubled CO<sub>2</sub> concentrations. *Plant. Cell. Environ.* 19, 585-592.
101. Oberbauer, S., Strain, B. R. and Fetcher, N., 1985, Effect of CO<sub>2</sub> enrichment on seedling physiology and growth of two tropical tree species. *Physiol. Plant*, 65: 352-356.
102. Oechel, W., and Strain, B.R. 1985. Native species responses to increased CO<sub>2</sub> In: Strain B.R. and Cure, J.D. (eds). *Direct effects of increasing CO<sub>2</sub> on vegetation*, pp. 117-154.
103. Patterson, D.T., Flint, E.P. and Beevens, J.L. 1984. Effects of CO<sub>2</sub> enrichment on competition between a C4 weed and a C3 crop. *Weed Sci.* 32, 101-105.

104. Peet, M.M., Huber, .C. and Patterson, D.T., 1986. Acclimation to high CO<sub>2</sub> in monaceous cucumbers. II. carbon exchange rates, enzyme activities and nutrient concentrations. *Plant Physiol.* 80, 63-67.
105. Picon, C., Guehl, J.M., Ferhi, A., 1996. Leaf gas exchange and carbon isotope composition responses to drought in a drought-avoiding ( *Pinus pinaster* ) and a drought -tolerant (*Quercus petraea*) species under present and elevated atmospheric CO<sub>2</sub> concentrations. *Plant. Cell. Environ.* 19, 182-190.
106. Poorter, H., Pat, S and Lambers, H. 1988. The effect of elevated atmospheric CO<sub>2</sub> concentration on growth, photosynthesis and respiration of *Plantago major* *Physiol. Plant.*, 73:553-559.
107. Potvin, C, and Strain, B.R. 1985. Photosynthetic responses- to growth temperature and CO<sub>2</sub> enrichment in two species- of C<sub>4</sub> plants. *Can. J. Bot.*, 63: 483-487.
108. Price, P.W., 1984. *Insect ecology* (2nd edn.). Wiley-Interscience, New York.
109. Radin, J.W., Kimball, B.A., Hendrix, D.L. and Mauny, J.R., 1987. Photosynthesis of cotton plants exposed to elevated levels of CO<sub>2</sub> in the field. *Photo. Res.*, 12: 191-203.
110. Raper, C.D.Jr., and Peedin, C.F. 1978. Photosynthetic rate during steady-state growth as influenced by CO<sub>2</sub> concentration. *Bot. Gazz.* 139, 147-149.
111. Rawland-Bamford, A.J., Baker, J.T., Allen, L.H. Jr., and Bowes, G. 1991. Acclimation of rice to changing atmospheric carbon dioxide concentration. *Plant Cell Environ.* 14, 577-583.
112. Reekie, E.G. and Bazzaz, F.A., 1989, Competition and pattern of resource use among seedlings of five tropical trees grown at ambient and elevated carbon dioxide. *Oecologia*, 79: 212-222.
113. Reuveni, J. and Gale, J. 1985. The effect of high levels of carbon dioxide on dark respiration and growth of plants. *Plant. Cell Environ.* 8:623-628.
114. Riviere, R.H., Contard, P. and Bestsche, T. 1996. Adaptation of pea to elevated atmospheric CO<sub>2</sub>: Rubisco, Phosphoenolpyruvate carboxylase and chloroplast phosphate translocator at different levels of nitrogen and phosphorus nutrition. *Plant. Cell. Environ.*, 19, 109-117.
115. Roden, J.S. and Ball M.C. 1996a. Growth and photosynthesis of two eucalyptus species during high temperature stress under ambient and elevated CO<sub>2</sub>. *Global Change Biol.* 2,115-128. ,
116. Roden J.S. and Ball M.C., 1996b. The effect of elevated CO<sub>2</sub> on growth and photosynthesis of two eucalyptus species exposed to high temperatures and water deficits. *Plant. Physiol.* 111, 909-919.



117. Rogers, H. H., Bingham, G.E., Cure, J. D., Smith, J. M. and Surano, K. A., 1983, Responses of selected plant species to elevated CO<sub>2</sub> in the field. *J. Environ. Qual.*, 12: 569.
118. Rogers, H.H., Cure, J.D. and Smith, J.M., 1986. Soybean growth and yield response to elevated CO<sub>2</sub>. *Agril. Eco. Environ.*, 16: 113-128.
119. Rogers, H.H., Cure, J.D., Thomas, J.F. and Smith, J.M., 1984. Influence of elevated CO<sub>2</sub> on growth of soybean plants. *Crop Sci.*, 24: 233-238.
120. Rogers, G.S., Milham P.J., Thibaud M.C. and Conroy J.P. 1996a. Interaction between rising CO<sub>2</sub> concentration and nitrogen supply in cotton I. Growth and leaf nitrogen concentration. *Aust. J. Plant. Physiol.* 23, 119-125.
121. Rogers, G.S., Milham, P.J., Gillings, M. and Conroy, J.P. 1996b. Sink strength may be the key to growth and nitrogen responses in N-dificient wheat at elevated CO<sub>2</sub>. *Aust. J. Plant. Physiol.* 23, 253-64
122. Ryle, G.J.A., Powell, C.E., and Tewson, V., 1992. Effect of elevated CO<sub>2</sub> on photosynthesis, respiration and growth on perennial rye grass. *J. Expt. Bot.*, 43: 811-818.
123. Sage, R.F. and Sharkey, T.D., 1989, Acclimation of photosynthesis to elevated CO<sub>2</sub> in five C<sub>3</sub> species. *Plant. Physiol.*, 89: 590-596.
124. Samarakoon, A.R. and Gifford, R.M., 1996a. Elevated CO<sub>2</sub> effects on water use and growth of maize in wet and drying soil. *Aust. J. Plant. Physiol.* 23, 53-62
125. Samarakoon, A.B. and Gifford, R.M., 1996b. Wateruse and growth of cotton in response to elevated CO<sub>2</sub> in wet and drying soil. *Aust. J. Plant. Physiol.* 23, 63-74.
126. Sasek, T.W. and Strain, B.R., 1989. Effects of CO<sub>2</sub> on the expansion and size of Kudzu (*Pueraria lobata*) leaves. *Weed Sci.*, 76: 23-28.
127. Scarscia-Mugnozza, G., De Angelis, P., Matteucci, G., and Valentini, R., 1996. Long tern exposure to elevated CO<sub>2</sub> in a natural quercus ilex L. community: net photosynthesis and photochemical efficiency of PS II at different levels of water stress. *Plant. Cell. Environ.* 19, 643-654.
128. Schenk, U., Manderscheid, R., Huguenot, J and Weigel, H.J. 1995. Effects of CO<sub>2</sub> enrichment and intraspecific composition on biomass partitioning, nitrogen content and microbial biomass carbon in soil of perennial regrass and white clover. *J.Expt. Bot.* 46 (2889) 987-993.
129. Schwanz P., Picon C., Vivin P., Dreyer E., 1996. Responses of antioxidative systems to drought stress in pedunculate oak and maritime pine as modulated by elevated CO<sub>2</sub>. *Plant. Physiol.* 110, 393-402.
130. Scriber, J.M. and Slansky, F. 1981. The nutritional ecology of immature insects. *Ann. Rev. Entomol.* 26, 183-211.

131. Schonfeld, M., Johnson, R.C. and Ferris, D.M., 1989. Development of winter wheat under increased atmospheric CO<sub>2</sub> and water limitations at tillering. *Crop Sci.*, 29: 1083-1086.
132. Senock, R.S., Ham, J.M., Loughin, T.M., Kimball, B.A., Hunsaker, D.J., Pinter, P.J., Wall, G.W., Gracia, R.L. and La Morte, 1996. Sap flow in wheat under free-air CO<sub>2</sub> enrichment. *Plant. Cell. Environ.* 19, 147-158.
133. Sharkey, T.D., 1985, Photosynthesis in intact leaves of C<sub>3</sub> plants : physics, physiology and rate limitations. *Bot. Rev.*, 51: 53-105.
134. Sharon, L.K. and Mitchell, C.A., 1988. Effects of CO<sub>2</sub> and photosynthetic photon flux on yield, gas exchange and growth rate of *Lactuca sativa* L. Cwaidmann's green, *J.Expt. Bot.* 39, 317-329.
135. Sheen, J. 1990. Metabolic repression of transcription in higher plants. *The plant Cell* 2: 1027-1038.
136. Sionit, N., Strain, B. R. and Besford, R. A., 1981, Environmental control on growth and yield of okra I. Effect of temperature and CO<sub>2</sub> enrichment at cool temperature. *Crop Sci.*, 21: 885-888.
137. Sionit, N., Strain, B. R., Hellmers, H. and Kramer, P. J., 1987, Effects of atmospheric CO<sub>2</sub> concentration and water stress on water relations of wheat. *Bot. Gazz.*, 142: 191-196.
138. Spencer, W. and Bowes, G., 1986, Photosynthesis and growth of water hyacinth under CO<sub>2</sub> enrichment. *Plant Physiol.*, 82: 528-533.
139. Stitt, M. 1991. Rising CO<sub>2</sub> levels and their potential significance for carbon flow in photosynthetic cells. *Plant Cell and Environ.*, 14: 741-762.
140. St. Omar, and Hovarth, S.M., 1983. Elevated carbon dioxide concentrations and whole plant senescence. *Ecol.* 64, 1311-1313.
141. Thomas, J.F. and Harvey, C.N., 1983. Leaf anatomy of four species grown under continuous CO<sub>2</sub> enrichment. *Bot. Gazz.*, 144: 303-309.
142. Thomas R.B. and Strain B.R. 1991. Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated CO<sub>2</sub>. *Plant Physiol.*, 96:627-634.
143. Thompson, G.B. and Drake, B.G., 1994. Insects and fungi on C3 sedge and a C4 grass exposed to elevated CO<sub>2</sub> concentrations in open-top chambers in the field. *Plant Cell Environ.* 17, 1161-1167.
144. Thompson, G.B., Brown, J.K.M., and Woodward, F.I. 1993. The effects of host carbon supply, nitrogen nutrition and water status on the infection of wheat by powdery mildew and aphids. *Plant Cell. Environ.* 16, 687-694.
145. Tissue, D.L., Thomas, R.B. and Strain B.R. 1993. Long-term effects of elevated CO<sub>2</sub> and nutrients on photosynthesis and Rubisco in loblolly pine seedlings. *Plant Cell. Environ.* 16, 859-865.

146. Tissue, D.L., and Oechel, W.C. 1987. Response of *Eriophorum vaginatum* to elevated CO<sub>2</sub> and temperature in the Alaskan tussock tundra. *Ecology* 68, 401-410.
147. Tolly, L. C. and Strain, B. R., 1985, Effects of CO<sub>2</sub> enrichment and water stress on gas exchange of *Liquidambar styraciflua* and *Pinus taeda* seedling grown under different irradiance. *Oecologia*, 65: 166-72.
148. Udayakumar, M. and T.G. Prasad. 1993. Assesment of limitations for photosynthetic rate in plants acclimated to elevated CO<sub>2</sub> and temperature. Proceedings of Indo-US workshop on Impact of Global Climate Changes on photosynthesis and plant productivity. pp.225-232.
149. Valle, R., Mishoe, J.W., Campbell, W. J., Jones, J. W. and Allen L. H. Jr., 1985, Photosynthetic response of Bragg soybean leaves adapted to different carbon dioxide environments. *Crop Sci.*, 25:333-339.
150. van Caemmerer S., and Farquhar, G.D. 1984. Effects of partial defoliation, changes in irritancy during growth, short term water stress and growth at enhanced p(CO<sub>2</sub>) on the photosynthetic capacity of leaves of *Phaseolus vulgaris* L. *Planta*. 160, 320-329.
151. Van ooster, J.J. and Besford, R.T., 1995. Some relationships between the gas exchange, biochemistry and molecular biology of photosynthesis during leaf development of tomato plants after transfer to different CO<sub>2</sub> concentration. *Plant. Cell. Environ.* 18, 1253- 1266.
152. Wang, Y.P. and Polglase, P.J. 1995. Carbon balance in the tundra forest during climate change : Scaling up from leaf physiology and soil carbon dynamics. *Plant. Cell. Environ.* 18, 1226-1244.
153. Waring, R.H. and Schlesinger, W.H. 1985. Forest ecosystem concepts and management. Academic press, inc. Orland, FL.
154. Wheeler, T.R., Morrison, J.I.L., Ellis, R.H. and Hadley, P. 1994. The effects of CO<sub>2</sub>, temperature and their interaction on the growth and yield of carrot (*Daucus carota*. L ). *Plant Cell Environ.* 17, 1275-84
155. Williams, M., Shewry, P.R., lawlor, D.W., and Harwood, J.L., 1995. The effects of elevated temperature and atmospheric carbon dioxide concentration on the quality of grain lipids in wheat (*Triticum aestivum* L.) grown at two levels of nitrogen application. *Plant. Cell. Environ.* 18, 999-1009.
156. Wilson, J.B., 1988, A review of evidence on the control of- shoot: root ratio, in relation to models. *Ann. Bot.* 61:433-439.
157. Woodrow, I.E. 1994. Optimal acclimation of the C<sub>3</sub> photosynthesis system under elevated CO<sub>2</sub>. *Photosynth. Res.* 39, 401-412.
158. Wong, S.C. 1990. Elevated atmospheric partial pressure of CO<sub>2</sub> and plant growth II Non-structural carbohydrate content in cotton plants and its effect on growth parameters. *Photo. Res.* 23,171-180.

159. Wong, S. C.,-1979, Elevated atmospheric partial pressure of CO<sub>2</sub> and plant growth, I. Interactions of nitrogen nutrition and photosynthesis capacity in C<sub>3</sub> and C<sub>4</sub>. *Oecologia*, 44: 68-74.
160. Wong, S. C. and Dunin, F. X., 1987, Photosynthesis and transpiration of trees in a Eucalyptus forest stand: Light and humidity responses. *Aust. J. Plant Physiol.*, 14: 619-632.
161. Wong, S.C. Kriedmann, P.E. and Farquhar, G.D. 1992. CO<sub>2</sub>xNitrogen interaction on seedling growth of four species of eucalyptus. *Aust. J. Bot.* 40, 457-472.
162. Wulff, R. and Strain, B. R., 1982, Effect of CO<sub>2</sub> enrichment on growth and photosynthesis in *Desmodium paniculatum*. *Can. J. Bot.*, 60: 1086-91.
163. Wullschlegel S.D., Norby, R.L., and Hendrix, D.L. 1992. Carbon exchange rates, chlorophyll content and carbohydrate status of two forest species exposed to CO<sub>2</sub> enrichment. *Tree Physiol.* 10, 21-31.
164. Yelle, S., Beeson, R.C. Jr., Trudel, M.J. and Gosselin, A. 1989. Acclimation of two species to high atmospheric CO<sub>2</sub>. I: Sugar and starch concentrations. *Plant Physiol.* 90, 1465-1472.
165. Zang H. and Nobel P.S., 1996. Photosynthesis and carbohydrate partitioning for the C3 desert shrub *Encelia farinosa* under current and doubled CO<sub>2</sub> concentrations. 110, 1361-1366.
166. Ziska, L.H. Drake, B.G. and Chamberlin, S., 1990. Long term photosynthetic response in single leaves of a C<sub>3</sub> and C<sub>4</sub> salt marsh species grown at elevated atmospheric CO<sub>2</sub> in situ., *Oecologia*, 483:469-472.
167. Ziska, L.H., Hogan, K.P., Smith, A.P. and Drake, B.G. 1991. Growth and photosynthetic response of nine tropical species with long exposure to elevated CO<sub>2</sub>. *Oecologia*:86:383-389.
168. Ziska, L.H., Weerakoon, W., Namuco, O.S. and Pamplona, R. 1996. The influence of nitrogen on the elevated CO<sub>2</sub> response in field grown rice. *Aust. J. Plant. Physiol.*, 23, 53-62.
169. Zangler, A.R., and Bazzaz, F.A. 1984. The response of plants to elevated CO<sub>2</sub> II. Competitive interaction among annual plants under varying light and nutrients. *Oecologia*, 62, 412-17.